

THYMUS CITRIODORUS AS A SOURCE OF ANTIOXIDANTS

O. Pereira^{1,2,3*}, A.M. Peres^{4,5}, M.R.M. Domingues⁶, S. M. Cardoso^{3,4}



¹DTDT, Escola Superior de Saúde, Instituto Politécnico de Bragança, Portugal; ²Departamento de Fisiologia e Farmacologia, Universidade de Salamanca, Espanha;

³CERNAS - Escola Superior Agrária, Instituto Politécnico de Coimbra, Portugal; ⁴CIMO - Escola Superior Agrária, Instituto Politécnico de Bragança, Portugal

⁵LSRE - Escola Superior Agrária, Instituto Politécnico de Bragança, Portugal; ⁶Centro de Espectrometria de Massa, Departamento de Química, Universidade de Aveiro, Portugal



*oliviapereira@ipb.pt



INTRODUCTION



Thymus species are well known as medicinal plants because of their biological and pharmacological properties, which include anti-asthmatic, anti-septic, antimicrobial and antioxidant [1]. It is believed that part of these beneficial effects are due to the volatile constituents of *Thymus*, and thus, their essential oil composition has been the focus of many investigations. In contrast, there is only a limited number of data on the composition of other bioactive phytochemicals of *Thymus* and their potential biological effects.

The present study aims to elucidate the phenolic composition of an ethanolic extract of *Thymus citriodorus*, as well as to determine its antioxidant capacity.

METHODS

The ethanolic extract was obtained by solubilisation of the defatted-dried plant with aqueous ethanol (80%) for twenty minutes, in a total number of five extractions.

The total phenolic compounds of the ethanolic extract were determined by an adaptation of the Folin-Ciocalteu procedure [2].

The phenolic characterization was performed by fractionation of the extract by reversed-phase HPLC and further analysis of the major phenolic compounds by ESI-MS and MSⁿ [3].

The HPLC analysis was performed on a RP-C18 column 250 mm×4 mm id, 5µm bead diameter (Temperature of 30°C, flow rate of 1 mL/min). Gradient elution was carried out with a mixture of 0.1% (v/v) of formic acid in water and acetonitrile and the chromatographic profiles were recorded at 280 nm.

The antioxidant activity was accessed by measuring the 2,2-diphenyl-1-picrylhydrazyl radical (DPPH) scavenging potential [4] and its reducing power [5].

RESULTS AND DISCUSSION

Table 1- Extraction yields, phenolic content and antioxidant capacity of *Thymus citriodorus*

Mass (% of dry weight)	Total Phenolics ^a (mg/g fraction)	DPPH (EC ₅₀) ^b (mg/mL)	Reducing Power (EC ₅₀) ^c (mg/mL)
17.1	139±14	0.32±0.05	0.8±0.2

Values are means ± S.D. of three replicate analyses;

^a Data expressed as milligrams of gallic acid equivalents (GAE) per gram of extract;

^b EC₅₀ – Concentration for a 50% inhibition;

^c EC₅₀ – Effective concentration at which the absorbance was 0.5.

The phenolic compounds account for 13.9% of the ethanolic extract total and this extract exhibited a high antioxidative capacity, with EC₅₀ values of 0.32±0.05 mg/ml and 0.8±0.2 mg/ml, for the DPPH scavenging potential and for the reducing power, respectively (Table 1).

The main phenolic components of the ethanolic extract of *Thymus citriodorus* were luteolin-7-*O*-glucoside (12±2 µg/mg extract), rosmarinic acid (10.4±0.6 µg/mg extract) and apigenin- 7-*O*- glucuronide (9±2 µg/mg extract) (Table 2).

CONCLUSION

- The ethanolic extract of *Thymus citriodorus* has a good antioxidant capacity.
- This extract is mostly rich in luteolin-7-*O*-glucoside, rosmarinic acid and apigenin- 7-*O*- glucuronide.
- Yet, it also contains phenolic compounds that were not previously found in *Thymus* genus.
- New compounds enclose glucosides of common flavonoids and sagerinic acid.
- The relevance of the main phenolic component in the beneficial properties of this plant is now under investigation.

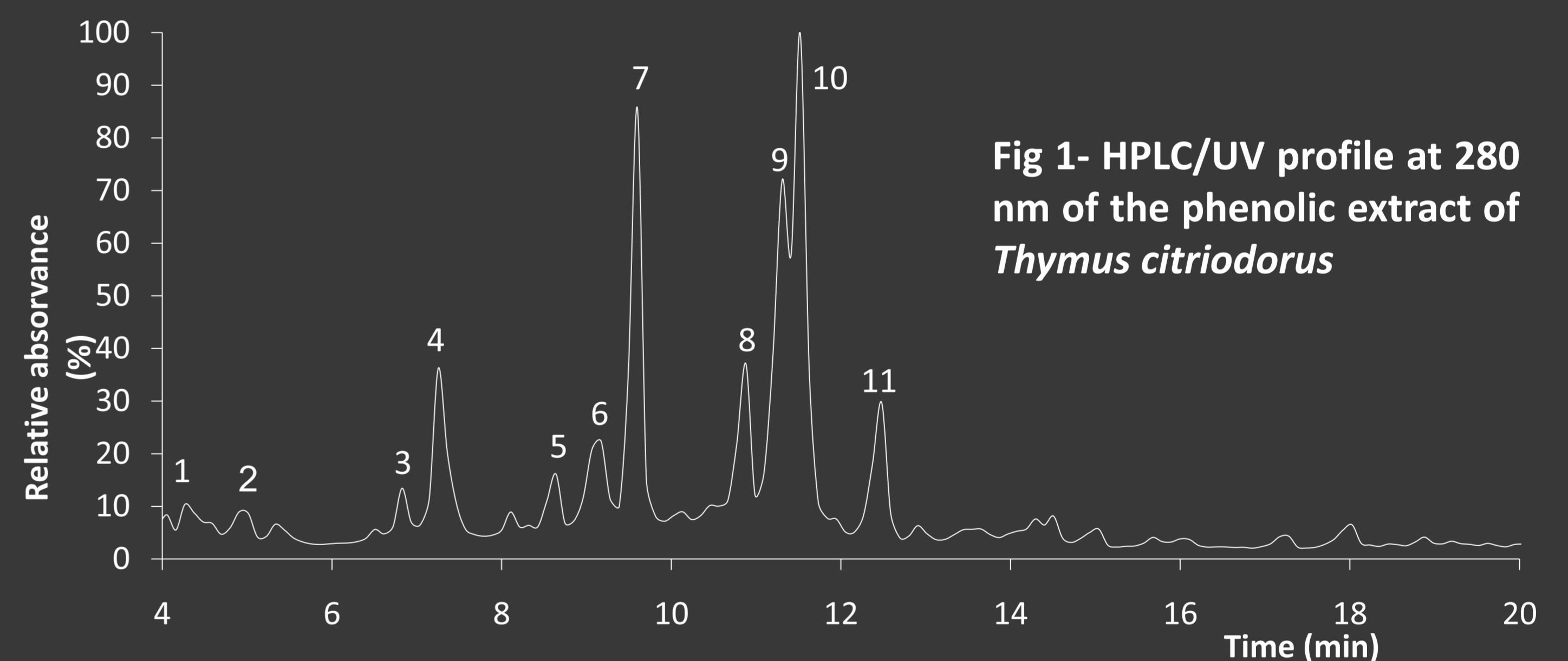


Fig 1- HPLC/UV profile at 280 nm of the phenolic extract of *Thymus citriodorus*

Table 2- Mass spectral data of the main phenolic constituents isolated from the *Thymus citriodorus*

Peak	RT (min)	Compound	Standard Compound	Mean content (mg/g extract)
1	4.3	5' hidroxijasmonic acid 5'- <i>O</i> -glucoside Eriodictyol-3',7-di- <i>O</i> -glucoside	E-7- <i>O</i> -G	0.71±0.07
2	5.0	Non identified	-	-
3	6.8	Eriodictyol- <i>O</i> -glucoside Quercetagenin-dimethyl-ether- <i>O</i> -hexoside	E-7- <i>O</i> -G	1.3±0.4
4	7.3	Eriodictyol- <i>O</i> -glucoside	E-7- <i>O</i> -G	3.7±0.5
5	8.6	Luteolin-5- <i>O</i> -glucoside	L-7- <i>O</i> -G	3.2±0.5
6	9.1	Naringenin-5- <i>O</i> -glucoside Eriodictyol- 7- <i>O</i> -glucuronil	N-7- <i>O</i> -G	1.8±0.2
7	9.6	Luteolin-7- <i>O</i> -glucoside Luteolin-7- <i>O</i> -glucuronil Sagerinic acid	L-7- <i>O</i> -G	12±2
8	10.9	Chrysoeriol-7- <i>O</i> -glucoside	-	-
9	11.3	Apigenin- 7- <i>O</i> - glucuronide	A-7- <i>O</i> -G	9±2
10	11.5	Rosmarinic acid	RA	10.4±0.6
11	12.5	3'- <i>O</i> -(8''- <i>Z</i> -Caffeoyl)rosmarinic acid	RA	2.3±0.9

E-7-*O*-G: Eriodictyol-7-*O*-glucoside; L-7-*O*-G: Luteolin-7-*O*-glucoside; N-7-*O*-G: Naringenin-7-*O*-glucoside; A-7-*O*-G: Apigenin-7-*O*-glucoside; RA: Rosmarinic Acid

REFERENCES

- [1] Bonanni, A. *et al.* (2007). *Food Chem.* 102, 751-8
- [2] Guyot, S. *et al.* (1998). *J. Agric. Food Chem.* 46, 1698-705
- [3] Falcão, S. *et al.* (2010). *Anal. Bioanal. Chem.* 396, 887-897
- [4] Ferreira, A. *et al.* (2006). *J. Ethnopharmacol.* 108, 31-7
- [5] Pereira, J. *et al.* (2006). *J. Agric. Food Chem.* 54, 8425-8431

ACKNOWLEDGEMENT

Olívia Pereira thanks for the PROTEC grant SFRH/PROTEC/49600/2009 (Programa de apoio à formação avançada de docentes do Ensino Superior Politécnico).