



## Review

## Phytochemical and bioactive potentials of African Annonaceae species

Josefa Rangel<sup>a,b,c,d,1</sup>, Ângela Liberal<sup>c,d,1</sup>, Sílvia Catarino<sup>e</sup>, José Carlos Costa<sup>a</sup>, Maria M. Romeiras<sup>a,e,\*</sup>, Ângela Fernandes<sup>c,d,\*\*</sup>

<sup>a</sup> Linking Landscape, Environment, Agriculture and Food Research Center (LEAF), Associated Laboratory TERRA, Instituto Superior de Agronomia (ISA), Universidade de Lisboa, Tapada da Ajuda, 1340-017 Lisboa, Portugal

<sup>b</sup> Centro de Botânica, Universidade Agostinho Neto, Avenida Ho Chi Minh, Prédio do CNIC, 1º andar, ala esquerda, Luanda, Angola

<sup>c</sup> Centro de Investigação de Montanhas, Instituto Politécnico de Bragança, Campus de Santa Apolónia, 5300-5253 Bragança, Portugal

<sup>d</sup> Laboratório Associado para a Sustentabilidade e Tecnologia em Regiões de Montanha (SusTEC), Instituto Politécnico de Bragança, Campus de Santa Apolónia, 5300-253 Bragança, Portugal

<sup>e</sup> Centre for Ecology, Evolution and Environmental Changes (cE3c) & CHANGE - Global Change and Sustainability Institute, Faculdade de Ciências, Universidade de Lisboa, Campo Grande, 1749-016 Lisboa, Portugal

## ARTICLE INFO

## Keywords:

Annonaceae

Traditional medicine

Africa

Nutritional value

Biochemical composition

## ABSTRACT

This review aims to gather available information on the medicinal, nutritional, and bioactive profiles of Annonaceae species in the African continent, sponsoring their use worldwide and mainly in African communities, where access to food and medicines for basic health care is scarce. >60 medicinal taxa were compiled, belonging to 22 genera, namely *Annickia*, *Annona*, *Anonidium*, *Artabotrys*, *Cleistochlamys*, *Cleistopholis*, *Dennettia*, *Duguetia*, *Greenwayodendron*, *Hexalobus*, *Isolona*, *Lettowianthus*, *Monanthotaxis*, *Monodora*, *Neostenanthera*, *Polyceratocarpus*, *Sphaerocoryne*, *Uvaria*, *Uvariastrum*, *Uvariadendron*, *Uvariopsis* and *Xylopia*; the most diverse and economically important genera were the genera *Annona*, *Uvaria* and *Xylopia* with 7 species each. Annonaceae species hold a valuable nutritional profile, rich in proteins, fibers, and minerals, being also good sources of a wide range of bioactive compounds of high biological relevance. These compounds are especially important in developing countries, where most of these species are available for direct use as food and/or medicines by the most deprived populations.

## 1. Introduction

The world's human population is increasing, and it is estimated that there will be about 10 billion people on the planet in 2050. Because of this exponential growth, greater food production is required to fulfil human needs (Goncharova & Merzlyakova, 2022). However, there is little room for cropland expansion, while water scarcity is threatening future agricultural production potential (Ehara et al., 2018).

Identifying wild plants, locally used for food and in traditional medicine, can contribute to improve the quality of life, food security, and basic health care, particularly in tropical dry regions (Sapit & Padulosi, 2012). Most of these plants can be grown on underutilized lands, and are tolerant to arid and adverse environmental conditions,

but are only of local or regional significance, being known as underutilized, neglected, under-exploited, under-valued, or orphan species (Okigbo & Anyaegbu, 2021). These species are important for the preservation of plant genetic resources, and their *ex-situ* and *in-situ* conservation is crucial to ensure their maintenance (Rocha et al., 2021). Despite the growing research interest in this field, and the fact that this type of species are important components of agrobiodiversity with the potential to adapt to climate change, they remain underutilized in conventional agriculture (Mabhaudhi et al., 2017).

Plant-based foods represent the main source of macro- and micro-nutrients, and essential minerals for human nutrition, holding bioactive properties that may contribute to the discovery of new medicines (Tekuri et al., 2019). This topic has been a major focus of attention

\* Correspondence to: M. M. Romeiras, Linking Landscape, Environment, Agriculture and Food Research Center (LEAF), Associated Laboratory TERRA, Instituto Superior de Agronomia (ISA), Universidade de Lisboa, Tapada da Ajuda, 1340-017 Lisboa, Portugal

\*\* Correspondence to: Â. Fernandes, Centro de Investigação de Montanhas, Instituto Politécnico de Bragança, Campus de Santa Apolónia, 5300-5253 Bragança, Portugal.

E-mail addresses: [mmromeiras@isa.ulisboa.pt](mailto:mmromeiras@isa.ulisboa.pt) (M.M. Romeiras), [afeitor@ipb.pt](mailto:afeitor@ipb.pt) (Â. Fernandes).

<sup>1</sup> These authors have the same contribution.

<https://doi.org/10.1016/j.foodchem.2024.139048>

Received 19 October 2023; Received in revised form 7 March 2024; Accepted 14 March 2024

Available online 21 March 2024

0308-8146/© 2024 Elsevier Ltd. All rights reserved.

worldwide, but the information available for several plant families, with distribution restricted to tropical regions, has widely been seen as modest in Africa (Catarino et al., 2021). Particularly, neglected, and underutilized species play a crucial role to fight against hunger and are a key resource for agriculture and rural development, with a high potential to contribute to global food security.

The plant family Annonaceae (the Soursop family) occurs in tropical to subtropical regions of all continents and comprises 122 genera and ca. 2440 species. Erkens et al. (2023) studied the distribution of this family and showed that it is present in four of the six main terrestrial biomes, distributed across about 109 countries, occurring mainly in the tropical region, in arboreal and other associated biomes; this means that their distribution is centred in the equatorial zone.

This family encompasses a wide variety of edible and non-edible plant species, extremely relevant at the medicinal and nutritional levels, providing edible fruits and allowing the extraction of oils from the seeds, leaves, and fruits of different species, able to be used as medicines, cosmetics, perfumes, and several other products (Handayani & Yuzammi, 2021).

Within this family, it outstands the genus *Annona* that has some species with horticultural and economic importance as fruit crops (Martin et al., 2019). It is known that Annonaceae species are valuable sources of bioactive compounds used mainly in traditional medicine by local communities to treat various diseases (Coria-Téllez et al., 2018).

Many of these species are odorous, due to the presence of essential oils which are mainly composed of mono- and sesquiterpene compounds (Fournier et al., 1999), and are used as biopesticides in the management of several insect pests (Maia et al., 2020). However, species widely used for food and medicines face greater extinction risk in their natural habitat, through combined over-exploitation, habitat loss and other factors (Pimenta et al., 2013).

Although the diversity of species with nutritional and pharmacological importance is recognized, there are still many knowledge gaps concerning this family, especially in African countries, where available information about the local flora is often scarce (Couvreur et al., 2006).

Approximately 40 genera and c. 500 species occur in Africa (Mols et al., 2004), where they are mainly found in tropical forests, but some have adapted to more arid climatic conditions and grow in savannahs (Couvreur, 2008). In this continent, and in addition to food and medicine, several Annonaceae species are used by local communities as timber to manufacture several materials and household and agricultural tools, to build houses and boats, to produce charcoal, and as firewood for cooking (Handayani, 2018).

In this study, we make a first attempt to review the available data on the African Annonaceae, namely their traditional medicinal uses, bioactive compounds, as well as nutritional and chemical composition, including mineral and vitamin contents, to promote their use across the world, and particularly by African communities, whose needs for food and medicines for basic health care need urgent solution.

### 1.1. Methods

We conducted a comprehensive review of the existing literature, following the typical guidelines of a review article. The aim of the review was to gather relevant and available information on African Annonaceae species, including their uses in traditional medicine, bioactive compounds, as well as their nutritional and chemical composition. To ensure a complete search, we used two electronic sources of scientific literature, namely Web of Science ([webofknowledge.com](http://www.webofknowledge.com)) and Scopus ([www.scopus.com](http://www.scopus.com)), from September 2022 to March 2023. Our search encompassed original articles, commentaries, books, letters, and reports related to the Annonaceae species most used in Africa.

We specifically focused on studies published between 1984 and 2023. The initial keywords used for the search were Annonaceae, Africa, main uses, nutritional and biochemical composition. These keywords were selected to identify literature that examines various aspects of the

family Annonaceae in general sense. We sought additional information from subject matter experts to refine our search strategy and locate additional relevant literature. With this, we were able to compile a final collection of 192 scientific studies for the analysis and synthesis of this review.

## 2. Traditional medicinal uses of Annonaceae species

In sub-Saharan African region, our investigation identified 65 Annonaceae species documented for their utilization in traditional medicine. Among these species, 63 are native, while two, namely *Annona muricata* and *Annona squamosa*, are naturalized. Introduced from America, these species are currently successfully established, capable of reproducing, and can disseminate spontaneously. Table S1 details the main ailments addressed, traditional preparation methods, modes of administration and the specific plant parts employed, such as leaves, fruits, seeds, and bark.

*Annona senegalensis* (Fig. 1) and *Xylopia aethiopica* stand out as the most reported species across distinct African countries, exhibiting diverse therapeutic applications. For example, in Cameroon, a decoction of *Annona senegalensis* bark is used for fever, respiratory infections, convulsions and snake bites (Tsabang et al., 2012). In the Democratic Republic of Congo (DRC), a decoction of the bark, leaves, and roots of the same species is employed for diabetes (Kasali et al., 2021), while in Guinea-Bissau, it is utilized for alleviating pains, addressing pregnancy complications, intestinal disorders, and skin inflammations (Catarino et al., 2016). *Xylopia aethiopica* seeds are applied for chest pains, coughs, and intestinal parasites in Angola (Costa & Pedro, 2013), while mashed fruits serve post-partum care in Cote d'Ivoire (Malan et al., 2015), and a decoction of dried fruits is used for aches and fever in Cameroon (Tsa-bang et al., 2012).

*Annona muricata*, *Duguetia staudtii*, and *Monodora myristica* (Fig. 1) are each documented in five countries. *Annona muricata* (Fig. 1) is predominantly utilized for its leaves and is documented for treating malaria in Angola, Cameroon, and Togo (Costa & Pedro, 2013; Koudouvo et al., 2011; Tsabang et al., 2012). In Guinea-Bissau and Nigeria, it is employed for addressing intestinal problems (Aiyelaja & Bello, 2006; Catarino et al., 2016). *Duguetia staudtii*, primarily used for its bark, is reported for malaria treatment in Cameroon and respiratory problems in Congo, Ghana, Cote d'Ivoire, and Nigeria (Sarpong et al., 2016; Tolu-Odugbemi, 2008; Tsabang et al., 2012). *Monodora myristica* is extensively used for its seeds, addressing diabetes in the DRC, pregnancy problems in Guinea-Bissau, and haemorrhoids in Cote d'Ivoire (Catarino et al., 2016; Malan et al., 2015).

Among the African countries, Cameroon leads with 34 reported Annonaceae medicinal species, followed by Nigeria (17) and Tanzania (9) (Fig. S1). Bark seems to be the most frequently used plant part (documented 51 times), followed by roots (39), leaves (31), and fruits (10). Decoction remains the preferred method of preparation, reported for 46 species.

Africa encompasses a rich diversity of cultures and indigenous knowledge systems that have long relied on plants for medicinal purposes, often utilizing the same plant species for various therapeutic applications across different regions (Table S1). Documenting these traditional uses of medicinal plants is critical for the preservation of valuable indigenous knowledge that may otherwise be lost in the near future (Romeiras et al., 2023). Moreover, understanding the diverse medicinal uses of Annonaceae species in Africa's developing countries, where the access to conventional medical services is limited, can inform healthcare practices, and contribute to improved public health outcomes.

In addition, the documentation of traditional uses of medicinal plants also enhances the potential for drug discovery, given that many modern pharmaceuticals are derived from natural compounds found in plants (Kayser, 2018). Therefore, investigating traditional medicinal applications not only offers guidance but also underscores opportunities



**Fig. 1.** Representative leaves, fruits, and seeds from different species. A - *Annona senegalensis* leaves; B - *A. senegalensis* fruits; C - *Monodora myristica* seeds; D - *A. muricata* fruits; E - *A. squamosa* fruits. (Photos by the author Josefa Rangel).

for pharmacological research.

### 3. Bioactive compounds in Annonaceae species

Plants are the main sources of bioactive compounds for the research of new drugs, producing substances of different chemical classes (Perone et al., 2022). Over the last few years, several studies have revealed the medicinal and phytochemical properties of Annonaceae species in Africa, Asia, and America; a wide range of bioactive compounds has been found, isolated, and characterized from different parts of the plants. Some of these compounds and their biological relevance for human health and well-being are presented in Table 1.

Phytochemical studies have revealed different classes of secondary metabolites, such as monoterpenes, diterpenes, triterpenes, flavonoids, phenylpropanoids, acetogenins, and, especially, isoquinoline derivatives. Some of these secondary metabolites isolated from Annonaceae species show important biological assets such as anti-inflammatory, trypanocide, leishmanicidal, antimalarial, antimicrobial, antioxidant, and antirheumatic activities, being also cytotoxic to human tumor cells. Liaw et al. (2010) identified acetogenins as the main class of compounds found in the Annonaceae family, these being exclusive to species native from tropical regions. The unique structure of this class of compounds (Parmena et al., 2012), makes acetogenins important natural compounds due to their antitumoral activity as well as for being potent antimalarial, immunosuppressive and pesticide agents (Keinan & Sinha, 2002).

Despite the effort to investigate the phytochemical and pharmacological properties of Annonaceae species in recent years, the number of studied species is still very low compared with their great biodiversity. According to Costa et al. (2021), only 15% of the Annonaceae species were subject to phytochemical and pharmacological investigation. Most of the studies focused on the genera *Annona*, *Asimina* and *Cananga*, and some species of the genera *Duguetia*, *Guatteria* and *Xylopia*, mainly due to their great economic relevance. In a study performed by Nyandoro et al. (2019), the crude ethanolic extract of the root bark of *Cleistochlamys kirkii* was able to inhibit 72% of the malaria parasite *Plasmodium falciparum*, this effect being attributed to the presence of cleistonol. Also, alkaloids and flavonoids isolated from the leaves of *Annona cherimola*

present antimicrobial, antiviral, and cytotoxic effects against the human prostate tumor cell line (PC-3), these biological activities being mainly due to cherimolacyclopeptide E, annomolin, and annocherimolin (Biba et al., 2014).

Anti-depressive effects have likewise been reported for the alkaloids annonaine, sinomenine, codeine, codeinone and hernagine in fruits of *Annona muricata* (Gavamukulya et al., 2017; Lee et al., 2021). In this species, toxicity against PC-3 prostate cancer cells was also reported, which is presumably due to the acetogenins annomuricin A, muricin J, muricin K, and muricin L present in its leaves, fruits, and pericarp (Sun et al., 2014). Acetogenins also seem responsible for the anticancer activity reported in *Uvaria acuminata* (Qian et al., 2015). Antioxidant, antidiabetic, hypolipidemic, hepatoprotective, antimicrobial, antiulcer, snake venom neutralizing and antiplasmodic activities have been reported in the leaves, roots, and fruits of *Uvaria chamae* (Ita, 2017; Monon et al., 2015).

In *Xylopia longipetala*, antimicrobial and antioxidant activities were also reported and associated with the occurrence of diterpenes  $\beta$ -pinene,  $\alpha$ -pinene, trans-pinocarveol, and myrtenol in the roots (Woguem et al., 2014). Biological and therapeutic effects such as anti-inflammatory, antispasmodic, diuretic, antihypertensive, cholesterol-lowering, antioxidant, antidiabetic, hepatoprotective, antibacterial, and antifungal, have been assigned to the occurrence of the monoterpenoids  $\alpha$ -phellandrene, p-cymene,  $\alpha$ -pinene, cis-sabinol, geraniol, eugenol camphene, myrcene, thujene, and limonene in *Monodora myristica* seeds (Agrimira & Siwela, 2017). Altogether, the available information shows not only the richness in bioactive compounds from different classes that can be found in several species of Annonaceae, but also their enormous biological and therapeutic relevance in traditional medicine and for the development of new drugs.

### 4. Nutritional and chemical composition of Annonaceae species

Several countries in tropical Africa have a wide variety of food sources that play a key role in human nutrition and well-being. Still, millions of people suffer from malnutrition, and most of the population depends on indigenous edible resources to meet their vitamin and mineral needs, which is why it is essential to sustainably exploit



**Table 1**

Bioactive compounds and their biological relevance in African Annonaceae species.

Scientific name	Used parts	Bioactivities	Type of study	Phenolic compounds or other isolated compounds	Therapeutic doses	Class of compounds	References
<i>Annickia chlorantha</i> (Oliv.) Setten & Maas	Stem barks	Antimalarial	<i>In vivo</i> (infected mice)	-	400 mg/kg (100% parasite clearance)	Alkaloids, saponins, phenolics, flavonoids and glycosides	(Adesokan & Akanji, 2010)
<i>Annona cherimola</i> Mill.	Leaves	Antioxidant Antiproliferative	<i>In vitro</i> HeLa and HepG2 cell lines	Anonaine, asimilobine, catechin, liriodenine, stepharine, lanuginosine, pronuciferine, and quercetin, kaempherol, luteolin, and apigenin derivatives	GI <sub>50</sub> 8.92 µg dw/mL cell medium (HeLa) GI <sub>50</sub> 10.22 µg dw/mL cell medium (HepG2)	Flavonoids Alkaloids	(Mannino et al., 2020)
<i>Annona glabra</i> L.	Fruits	Anti-inflammatory (inhibition of cyclooxygenase-2)	<i>In vitro</i> (Raw 264.7 mouse leukemic monocyte-macrophages)	Isodesacetylurvaricin	5 µM (barely detectable levels of COX-2 mRNA)	Acetogenins	(Wu et al., 2012)
<i>Annona muricata</i> L.	Leaves	Cytotoxic	<i>In vitro</i> : human breast carcinoma (MDA-MB-435S), human immortalized keratinocyte (HaCaT), and normal human hepatic cells (WRL-68)	Quercetin, robinetin, gallic acid, apigenin, luteolin, vitexin	Normal cells -IC <sub>50</sub> = 52.4 µg; Cancerous cells - IC <sub>50</sub> = 29.2 µg (MDA-MB-435S) and 30.1 µg (HaCaT)	Flavonols Polyphenols Flavones	(George et al., 2012; Jiménez et al., 2014; Menezes et al., 2019)
	Fruits (pulp)	Antioxidant		Cinnamic acid derivatives, coumaric acid, caffeoylquinic acid, and caffeic acid derivatives	-		
	Seeds	Antioxidant		Gallic acid, catechin, chlorogenic acid, caffeic acid, vanillin, coumaric acid, ferulic acid, quercetin, cinnamic acid, rutin	100 mg/mL (77.34% antioxidant activity)		
	Barks	Analgesic and anti-inflammatory		Kaur-16-en-19-oic acid	10 and 20 mg/kg	Ent-kaurane diterpenoid	
<i>Annona reticulata</i> L.	Leaves	Thrombolytic activity	<i>In vitro</i>	-	Methanol, ethanol and chloroform extract showed 68.81%, 72.06% and 78.17% inhibition of RBC hemolysis	Alkaloids, steroids, polyphenols	(Aggarwal et al., 2018; Shahriar et al., 2016)
	Fruits	Antioxidant, antibacterial, antifungal, antitumor, antidiabetic, hypocholesterolemic, antiatherosclerotic, insecticidal and larvicidal	<i>In vitro</i> human erythrocytes	<i>n</i> -hexadecanoic, octadecadienoic, pimarinic, and carboxylic acids, naphthalenemethanol, decahydro-5-(5-hydroxy-3-methyl-3-pentenyl)-1,4a-dimethyl-6-methylene- and spathulenol.	-	Polyphenols	
<i>Annona senegalensis</i> Pers.	Seeds	Anti-inflammatory	<i>In vivo</i> : LPS stimulated J774.2 mouse macrophages	Tryptamide alkaloid N-cerotoyltryptamine, acetogenin asimicin, kauranoic acid methylester, lacceroic acid, and stigmasterol glycoside	Plant extract - IC <sub>50</sub> = 8.7 ± 10.2 mg/mL  N-cerotoyltryptamine (isolated from the plant) - IC <sub>50</sub> = (2.7 ± 0.1 mg/mL)	Alkaloid, acetogenin	(Tamfu et al., 2021)
<i>Annona squamosa</i> L.	Leaves	Antiproliferative and citotoxic	<i>In vivo</i> : rats with breast cancer cell lines (MCF-7 and MDA-MB-231)	Germacrene-D, humulene, phytol	100 µg/mL	Polyphenols Flavonoids	(Adesanwo et al., 2020; Al-Nemari et al., 2022)
	Seeds Fruits	Antimicrobial Antioxidant	<i>In vitro</i>	Octadec-9-enoic acid, 9,10-dehydroisolongifolene and androsterone	Methanol extract – MIC = 0.39 mg/mL n-hexane fraction MIC = 1.56 mg/mL Antioxidant activity IC <sub>50</sub> = 1.33	Polyphenols	

(continued on next page)

Table 1 (continued)

Scientific name	Used parts	Bioactivities	Type of study	Phenolic compounds or other isolated compounds	Therapeutic doses	Class of compounds	References
<i>Artabotrys modestus</i> Diels	Bark	Antiviral	<i>In ovo</i>	Quebrachitol, cyclohexane-1,2,4,5-tetrol	360 µg/m	Alicyclic polyhydroxy compounds	(Nyandoro, 2017)
<i>Cleistochlamys kirkii</i> (Benth.) Oliv.	Bark	Antibacterial	<i>In vitro</i>	Dichamanetin, chamanetin, isochamanetin, α,β-unsaturated lactone (–)-cleistenolide	Dichamanetin – MIC = 1–7.5 µg/mL	Flavanones	(Nyandoro et al., 2019; Pereira et al., 2016)
	Bark, stem	Antimalarial and anticancer	<i>In vitro</i> - MDA-MB-231, triple-negative, aggressive breast cancer cell line	Dichamanetin, (E)-acetylmelodrinol, and cleistenolide	Antimalarial 0.01 µg/mL - 72% inhibition of <i>P. falciparum</i> <u>Anticancer</u> IC <sub>50</sub> = 42.0 µg/mL (crude extract) and 9.6–30.7 µM (isolated compounds)	Sesquiterpene	
<i>Cleistopholis patens</i> (Benth.) Engl. & Diels	Barks	Antibacterial, antioxidant	<i>In vitro</i>	–	100 mg/mL – 40% reducing ability 100 mg/mL against <i>Proteus vulgaris</i> and <i>Yersinia</i> ; 18 mg/ml against <i>Klebsiella aerogenes</i>	Aminoacids, amides, amines, carboxylic acid, carbonyl compounds, organic hydrocarbons, halogens, alcohols, phenols, sulfonamides	(Daniels & Ibiyemi, 2021)
<i>Duguetia staudtii</i> (Engl. & Diels) Chatrou	Barks	Anti-inflammatory, urease activity	<i>In vitro</i>	Pachypolignan, pachyophyllin, pachypodol, kumatakenin, etramethoxyflavone, benzopyran, corypalmine, polycarpol	Anti-inflammatory IC <sub>50</sub> = 3.89 to 14.13 µg/mL Urease activity IC <sub>50</sub> = 10.9 to 20.2 µg/mL Root bark extract - IC <sub>50</sub> 0.26 µg/mL; stem bark alkaloid fraction = IC <sub>50</sub> 0.27 µg/mL; root bark alkaloid fraction - IC <sub>50</sub> < 1 µg/mL; methanol fractions - 0.36 µg/mL to 0.69 µg/mL	Bisnorlignan, flavonoids, alkaloid, triterpenoid	
<i>Greenwayodendron suaveolens</i> (Engl. & Diels) Verdc.	Fruits, Leaves, Bark, Stem	Antiprotozoal	<i>In vitro</i>	Polycarpol and, dihydropolycarpol; polyalthenol and N-acetylpolyveoline	Crude methanol extracts of root bark - IC <sub>50</sub> = 1.97 µg/ml and leaves - IC <sub>50</sub> = 2.65 µg/ml	Triterpenes, Alkaloids	(Muganza et al., 2016)
<i>Isolona hexaloba</i> (Pierre) Engl. & Diels	Leaves, root barks, stems barks	Antiprotozoal	<i>In vitro</i>	–	Crude methanol extracts of root bark - IC <sub>50</sub> = 1.97 µg/ml and leaves - IC <sub>50</sub> = 2.65 µg/ml	Saponins, alkaloids, flavonoids, anthraquinones, steroids, tepernoids, coumarins	(Muganza et al., 2016)
<i>Monanthotaxis caffra</i> (Sond.) Verdc.	Leaves	Antihepatotoxic	<i>In vivo</i> (rats)	Crotopoxide, 5,6-diacetoxy-1-benzoiloximetil-1, 3-ciclohexadieno	100 mg/kg	Terpenes	(Makhuvele et al., 2022)
<i>Monanthotaxis discolor</i> (Diels) Verdc.	Root barks	Antimicrobial	<i>In vitro</i>	α-cadinol, (–)-alloaromadendrene, aristolone, γ-cadinene, δ-cadinene, cubenene	Dichloromethane extract - LC <sub>50</sub> 41.794 µg/ml Methanol extract - LC <sub>50</sub> 13.560 µg/ml Inhibitory activity against xanthine oxidase and urease - IC <sub>50</sub> 0.56 ± 1.72 and 17.86 ± 1.13 µg/mL, respectively; DPPH and Fe <sup>2+</sup> chelation -IC <sub>50</sub> 9.39 ± 0.39 and 14.91 ± 0.82 µg/mL, respectively	Sesquiterpenes	(Parmena et al., 2012)
<i>Monodora myristica</i> (Gaertn.) Dunal	Seeds	Anti-nephrolithiasis Antioxidant	<i>In vitro</i>	Gallic, chlorogenic, caffeic, p-coumaric, ellagic acids, catechin, rutin, quercetin and luteolin		Phenolic compounds	(Itrondi et al., 2023)
<i>Uvaria acuminata</i> Oliv.	All plant parts	Antimicrobial	<i>In vitro</i>	Diuvaretin, triuvaretin, angoluvarin	–	Dihydrochalcones	(Munissi, 2019)

(continued on next page)

Table 1 (continued)

Scientific name	Used parts	Bioactivities	Type of study	Phenolic compounds or other isolated compounds	Therapeutic doses	Class of compounds	References
<i>Uvaria afzelii</i> Scott Elliot	Roots	Hepatoprotective effect	<i>In vivo</i> (rats)	Syncarpic acid, dimethoxymatteucinol, emorydone, uvafzelic acid, syncarpurea, afzeliindanone	125, 250 and 500 mg/kg conjugated with CCl <sub>4</sub>	–	(Ofeimun et al., 2013)
<i>Uvaria angolensis</i> Welw. ex Oliv.	Leaves, twigs and stem bark	Antiplasmodial	<i>In vivo</i> (albino mice) <i>In vitro</i>	–	400 mg/kg throughout 4 days	Acetogenins, alkaloids, phenols, lactones	(Mfopa et al., 2017)
<i>Uvaria chamae</i> P. Beauv.	Roots	Antioxidant Antibacterial	<i>In vitro</i>	–	Ethanol extract – MIC = 2 Aqueous extract – MIC = 16 DPPH – IC <sub>50</sub> = 3.52 to 14.35 µg/mL	Tannins, flavonoids, and phenols Acetogenins	(Monon et al., 2015) (Fall et al., 2006; Jalil et al., 2020)
	Seeds	Anti-inflammatory		Joolanin, squamocin, desacetylurvaricin, chamuvarinin			
<i>Uvaria lucida</i> Bojer ex Benth.	Root bark	Antimalarial	–	Lucidene, uvaletin, diuvaretin, chamuvaretin	–	Sesquiterpene, dihydrochalcones	(Weenen et al., 1990)
<i>Uvaria pandensis</i> Verdc.	Leaves	Antibacterial, cytotoxic (human breast cancer)	<i>In vitro</i>	(–) -Pipoxide, (+) -Pandoxide, pandensinol D - F	Antibacterial (8'α,9'β-dihydroxy)-3-farnesylindole - EC <sub>50</sub> = 9.8 µM against <i>B. subtilis</i> Cytotoxicity EC <sub>50</sub> > 100 µM	Cyclohexene derivatives, flavonoids	(Maeda et al., 2022)
<i>Uvaria tanzaniae</i> Verdc.	Stem and root bark	Antimalarial	–	Tanzanene	–	Sesquiterpene	(Weenen et al., 1991)
<i>Uvaria versicolor</i> Pierre ex Engl. & Diels	Stems	Acaricidal	<i>In vitro</i>	Benzyl benzoate, versuvanone, oxoaporphine, liriodenine	Methanol and hexane extracts with EC <sub>50</sub> values of 0.095 g/m <sup>2</sup> and 0.12 g/m <sup>2</sup> , respectively	Flavanone	(Akendengue et al., 2003)
<i>Uvariadendron angustifolium</i> (Engl. & Diels) R. E.Fr.	Stem, stem bark, leaves, and root	Cytotoxic activity on MCF-7 human breast cancer cells	<i>In vitro</i>	Neral, geranial, methyl eugenol,	IC <sub>50</sub> values of the leaf (220 µg/mL), stem (270 µg/mL), stem-bark (320 µg/mL) and root (460 µg/mL) volatile extracts µg/mL	Monoterpenes	(Noudogbessi et al., 2014)
<i>Xylopia hypolampra</i> Mildbr.	Bark, stem	Antimicrobial	<i>In vitro</i>	Verbenone, borneol, eucalyptol, nopinone, <i>trans</i> -pinocarveol, α-terpineol, <i>para</i> -cymen-8-ol, terpinen-4-ol, cyperotundone, myrtenal	MIC >500 µg/mL	Terpenes, monoterpenes, sesquiterpenes	(Pedrali et al., 2019)
<i>Xylopia longipetala</i> De Wild. & T. Durand	Fruits	Chemopreventive, anti-inflammatory, antimicrobial	<i>In vitro</i>	β-pinene, α-pinene, <i>trans</i> -pinocarveol, myrtenol	IC <sub>50</sub> = 6.56 µg/ml (MDA-MB 231); 7.47 µg/ml (A375); 6.63 µg/ml (HCT116)	Monoterpene hydrocarbons Alkaloids	(Nishiyama et al., 2004; Woguem et al., 2014)
	Bark, roots	Analgesic and antiplasmodic	–	Xylopinidine, dehydrocoreximine, <i>N,N</i> -dimethylanomurine, methylphoebine	–		
<i>Xylopia staudtii</i> Engl. & Diels	Bark	Antimicrobial	<i>In vivo</i> (immunodepressed mice)	Kaurenoic acid	Bacterial load in faeces reduced 100% with 400 mg/kg	Diterpenes	(Nguiam et al., 2021)

available local resources. Particularly, in rural areas, malnutrition can be overcome through the consumption of local unknown products which might be rich sources of proteins, carbohydrates, and essential vitamins (Atasie et al., 2009). An example of this are some wild plants, whose fresh leaves, seeds, fruits, among other parts, can serve as the basis for a more balanced diet, especially among the poorest communities (Ruffo et al., 2002) and at times of seasonal food shortages (Umaru et al.,

2007).

The fruits of some Annonaceae species are mainly consumed *in natura* but can also be processed into different food products such as ice cream, juices, flakes, jellies, nectars, purees, yoghurts, syrups, alcoholic beverages, and extracts with medicinal properties (Fuentes et al., 2022). Table 2 presents the available data on the nutritional and energetic composition, particularly of fruits of Annonaceae species native to

**Table 2**

Nutritional composition (g/100 g dw) and energetic value (g/100 kcal dw) of African Annonaceae species.

Scientific name	Plant Parts	Moisture (% fw)	Ash (g/100 g dw)	Crude protein (g/100 g dw)	Crude Fiber (g/100 g dw)	Fat (g/100 g dw)	Carbohydrate (g/100 g dw)	Energy (kcal/100 g dw)	References
<i>Annickia chlorantha</i> (Oliv.) Setten & Maas	Stem bark	3.85 ± 0.35	2.5 ± 0.1	10.8 ± 0.2	72.3 ± 0.4	3.8 ± 0.2	6.3 ± 0.2	–	(Dawodu et al., 2014)
<i>Annona cherimola</i> Mill.	Pulp	–	0.9 ± 0.17	1.5 ± 0.2	–	–	–	–	(Cordeiro et al., 2013)
<i>Annona muricata</i> L.	Pulp	0.018 ± 0.001	0.006 ± 0.001	1.38 ± 0.01	3.02 ± 0.01	9.75 ± 0.01	67.14 ± 0.09	364.5	(Akomolafe & Ajayi, 2015; Coria-Téllez et al., 2018; Badrie & Schauss, 2010)
	Leaves	4.95	7.5	11.4	5.4	0.2	75.7	35.0	
	Fruit	82.8	60	1.0	0.8	0.97	14.6	53.1–61.3	
<i>Annona senegalensis</i> Pers.	Seeds	12.2	12.1	8.80	17.6	24.0	25.3	–	(Yisa et al., 2010)
<i>Annona squamosa</i> L.	Pulp and Fruits	78.54 ± 0.14	2.8 ± 0.2	1.13 ± 0.01	5.90 ± 0.05	0.79 ± 0.03	10.80 ± 0.06	51.8 ± 0.3	(Abdualrahman et al., 2019)
	Seeds	6.65 ± 0.05	5.24 ± 0.04	0.03	17.6 ± 0.1	17.6 ± 0.1	21.8 ± 0.1	434.5 ± 0.2	
<i>Cleistopholis glauca</i> Pierre ex Engl. & Diels	Fruits	–	1.2	5.0	29.7	–	9.7	57.3	(Lamperti et al., 2014)
<i>Duguetia staudtii</i> (Engl. & Diels) Chatrou	Fruits	–	1.8	5.4	53.8	–	19.8	–	(Lamperti et al., 2014)
<i>Greenwayodendron suaveolens</i> (Engl. & Diels) Verdc.	Young leaves	9.98 ± 0.29	6.14 ± 0.01	01.34 ± 0.01	05.6 ± 0.1	14.8 ± 0.1	72.3 ± 0.02	–	(Oshomoh et al., 2022)
<i>Monodora myristica</i> (Gaertn.) Dunal	Seeds	14.6 ± 0.4	2.4 ± 0.2	14.0 ± 0.8	21.0 ± 1.1	31.0 ± 0.5	16.9	402.8	(Borquaye et al., 2017)
<i>Monodora tenuifolia</i> Benth.	Seeds	19.0 ± 0.1	4.9 ± 0.6	8.3 ± 1.0	26.2 ± 1.2	34.9 ± 1.9	6.6 ± 0.7	–	(Chukwuma & Uzoma, 2013)
			5.1 ± 0.5			6.5 ± 0.8			
<i>Xylopia aethiopica</i> (Dunal) A.Rich.	Pulp	12.3 ± 0.2	0.5		18.3 ± 0.9	0.8	50.1 ± 0.5	289.6	(Borquaye et al., 2017; Fategbe et al., 2021)
	Whole fruit	11.3 ± 0.1	10.68 ± 0.02	7.7 ± 0.9	18.4 ± 0.4	32.7 ± 0.1	30.4 ± 0.4	–	
	Seeds	9.70 ± 0.05	6.60 ± 0.04		4.61 ± 0.04	0.1	54.52 ± 0.02	–	
<i>Xylopia longipetala</i> De Wild. & T.Durand	Seeds	8.95 ± 1.13	6.3 ± 0.4	8.3 ± 0.2	8.3 ± 0.6	6.1 ± 0.5	62.5 ± 0.8	80.8 ± 0.3	(Yanda-Shaba, 2017)
<i>Xylopia rubescens</i> Oliv.	Fruits	–	2.2	4.9	52.4	–	3.9	–	(Lamperti et al., 2014)
<i>Xylopia staudtii</i> Engl. & Diels	Fruits	–	1.6	8.6	43.7	21.4	6.5	–	(Lamperti et al., 2014)

Africa. Low-fat concentrations were found in the fruits of some of them, with *Annona muricata* leaves (0.2 g/100 g dw) and *Annona squamosa* fruits (0.79 g/100 g dw, Fig. 1) exhibiting the lowest values (López-Martínez et al., 2022). On the other hand, highest fat concentrations were found on seeds from *A. senegalensis* Pers (24.0 g/100 g dw) (Yisa et al., 2010), *A. squamosa* L. (17.6 g/100 g dw) (Abdualrahman et al., 2019), *M. myristica* (31.0 g/100 g dw) (Abdualrahman et al., 2019), *M. tenuifolia* Benth (34.9 g/100 g dw) (Chukwuma & Uzoma, 2013), and *X. aethiopica* (28.6 g/100 g dw) (Borquaye et al., 2017; Fategbe et al., 2021), as well as in the fruits of *X. staudtii* and *X. aethiopica* (21.4 and 32.7 g/100 g dw, respectively) (Lamperti et al., 2014), and young leaves of *G. suaveolens* (14.8 g/100 g dw) (Oshomoh et al., 2022), these parameter being highly influenced not only by the species, but also by the different parts of the plant used, along with other genetic and edaphoclimatic factors.

Annonaceae species are also characterized by low protein concentrations, the highest values being found in *Annona squamosa* seeds (18.34 g/100 g dw) (Abdualrahman et al., 2019). The highest moisture contents were identified in *Annona* genus, with the highest levels being identified in *Annona muricata* (82.8%) (Akomolafe & Ajayi, 2015) and in *Annona squamosa* (78.5%) fruits (Abdualrahman et al., 2019). The ash content, representing the mineral amounts present in each part of the plant (Imo et al., 2018), seems to vary between species, with *Annona senegalensis* presenting the highest concentration (12.1 g/100 g dw) and *Annona muricata* (0.006 g/100 g dw) the lowest (Akomolafe & Ajayi, 2015).

In general, despite their interesting nutritional profile, the proximate composition of Annonaceae fruits may considerably vary depending on species, soil characteristics, harvesting time, genetics, environmental conditions (i.e., temperature and precipitation), among other factors.

Also, the recorded variation could be attributed to the use of different methodologies to quantify the nutritional parameters.

Plants are one of the main sources of vitamins and minerals necessary for the proper functioning of living organisms, playing, among others, a key role in resistance against pathogens (Yanda-Shaba, 2017). However, many of such plants are often unknown, underutilized, and seasonal (Ayessou et al., 2009), factors that may restrict their everyday use. Some of these micronutrients are essential minerals, such as potassium (K), iron (Fe), magnesium (Mg), zinc (Zn), important for both human nutrition and prevention of various chronic and acute diseases that may be associated with imbalances of one or more of these minerals (El-Ramady et al., 2022).

The necessary concentration of minerals to conduct certain biological functions is variable, and some may be present in smaller amounts and act concomitantly with others. For example, lower levels of zinc can support calcium functions, as zinc helps calcium absorption into bones and, in turn, during some metabolic processes, certain enzymes depend on zinc to perform their vital functions (Imo et al., 2018).

In Table 3, the mineral profiles of some species of the Annonaceae family are presented. *Annona muricata*, for example, presents high levels of calcium in its fruit pulp (857.2 mg/100 g), and elevated levels of phosphorus (146.3 mg/100 g) have been reported in its seeds (Chimbevo & Essuman, 2019). High potassium levels were reported for the fruits of *Monodora myristica* (869.64 mg/100 g dw) (Raphael et al., 2010), *Annona cherimola* (406.8 mg/100 g dw) (Cordeiro et al., 2013), and *Xylopia aethiopica* (288.7 mg/100 g), the latter also with interesting amounts of magnesium (175.3 mg/100 g) (Fategbe et al., 2021). High amounts of calcium were found in *Annona muricata* pulp (857.2 mg/100 g) and in the seeds of *Annona squamosa* (648.5 mg/100 g) (Chimbevo & Essuman, 2019).

**Table 3**  
Mineral composition (mg/100 g dw) of African Annonaceae species.

Scientific name	Plant Part	Calcium	Sodium	Potassium	Phosphorus	Zinc	Iron	Magnesium	Copper	References
<i>Annickia chlorantha</i> (Oliv.) Setten & Maas	Fruits	0.36	0.06	–	0.13	17.8	104	0.28	27.2	(Lamperti et al., 2014)
<i>Annona cherimola</i> Mill.	Fruit	11.3	15.9	406.8	17.9	0.13	0.12	177.0	0.06	(Cordeiro et al., 2013)
	Pulp	857.2 ± 6.4	843.4 ± 16.3	322.3 ± 13.1	32.5 ± 0.9	0.40 ± 0.03	1.05 ± 0.06	24.0 ± 0.8	1.00 ± 0.03	(Chimbevo & Essuman, 2019)
<i>Annona muricata</i> L.	Seeds	158.4 ± 2.9	21.0 ± 1.4	354.6 ± 2.2	146.3 ± 4.0	0.46 ± 0.04	3.6 ± 0.1	13.7 ± 0.3	0.04 ± 0.01	(Yisa et al., 2010)
<i>Annona senegalensis</i> Pers.	Seeds	135	–	47	–	48	180	24	29	(Yisa et al., 2010)
	Pulp	455.0 ± 2.9	10.4 ± 0.4	48.0 ± 0.6	28.7 ± 1.0	0.32 ± 0.02	1.65 ± 0.04	395.5 ± 4.6	0.10 ± 0.01	(Chimbevo & Essuman, 2019)
<i>Annona squamosa</i> L.	Seeds	648.5 ± 2.5	31.4 ± 0.6	23.3 ± 0.7	22.2 ± 0.8	0.31 ± 0.02	2.07 ± 0.02	52.0 ± 1.0	0.03 ± 0.01	(Lamperti et al., 2014)
<i>Cleistopholis glauca</i> Pierre ex Engl. & Diels	Fruits	140	40	–	80	11.7	118	80	8.5	(Lamperti et al., 2014)
<i>Cleistopholis patens</i> (Benth.) Engl. & Diels	Fruits	340	40	–	90	16.9	50.5	17	7.7	(Lamperti et al., 2014)
<i>Duguetia staudtii</i> (Engl. & Diels) Chatrou	Fruits	170	20	–	80	8.6	40.5	70	7.9	(Lamperti et al., 2014)
<i>Monodora myristica</i> (Gaertn.) Dunal	Seeds	416.0 ± 1.4	17.7 ± 0.3	869.6 ± 4.0	112.0 ± 4.6	1.5 ± 0.1	21.7 ± 0.5	87.0 ± 4.0	0.19 ± 0.02	(Raphael et al., 2010)
<i>Uvaria chamae</i> P.Beauv.	Stem bark	34.5 ± 0.9	2.7 ± 0.6	4.2 ± 0.2	1.8 ± 1.0	0.8 ± 0.4	3.4 ± 0.7	1.3 ± 0.5	–	(Ojokuku et al., 2011)
<i>Xylopia aethiopica</i> (Dunal) A.Rich.	Whole fruit	236.4 ± 2.0	15.77 ± 0.08	288.7 ± 0.1	62.75 ± 0.05	4.14 ± 0.04	32.5 ± 0.5	175.3 ± 1.2	1.55 ± 0.02	(Fategbe et al., 2021)
	Seeds	212.2 ± 1.2	13.60 ± 0.04	246.9 ± 0.6	59.0 ± 1.1	2.8 ± 0.6	29.0 ± 0.4	168.2 ± 0.7	1.32 ± 0.02	(Lamperti et al., 2014)
<i>Xylopia rubescens</i> Oliv.	Fruits	120	10	–	50	47.4	111	50	22.5	(Lamperti et al., 2014)

In addition to their mineral contents, Annonaceae species are rich in vitamins, such as vitamins C, E, thiamin, riboflavin, niacin, among others (Fuentes et al., 2022), which may provide several health benefits for living organisms (Scrimshaw, 2007). According to Floegel et al. (2011), some of these nutraceuticals act as antioxidants to prevent various acute and chronic diseases. Likewise, vitamins A, B, and C are non-enzymatic antioxidants and dietary supplements needed for human health, these being able to prevent and/or stop the development of some diseases, such as cancer, cardiovascular, and inflammatory diseases, among other health conditions (Didier et al., 2023).

In Table 4, the vitamin profile of some species of the Annonaceae family are presented; among them, *Annona squamosa* pulp presents the highest amount of vitamin C (25.6–58.8 mg/100 g dw) (Fuentes et al., 2022). Overall, this is the vitamin present in greatest amounts in all the analyzed species, with also interesting contents in *Annona cherimola* fruits (12.6–25.4 mg/100 g) and in *Monodora myristica* seeds (9.9 mg/100 g dw) (Nkwocha et al., 2019).

In addition to their richness in bioactive compounds, Annonaceae species are also excellent sources of other compounds (Ben-Othman et al., 2020; Bhardwaj et al., 2019), such as sugars, organic acids, essential aminoacids, and fatty acids, which can be found in leaves, seeds, roots, and stems (Nam et al., 2018). Organic acids play a fundamental role in plants, as they are responsible for their flavor and texture, thus promoting their acceptance in the market (Arena et al., 2021).

There are very few studies reporting the occurrence of organic acids in Annonaceae species. In *Annona cherimola*, for example, several organic acids had been identified, namely tartaric, malic, ascorbic, succinic, lactic, acetic, citric, and oxalic acids, while in *Annona reticulata* only ascorbic and oxalic acids were reported (González-Agüero et al., 2016; Kn et al., 2016).

The composition of sugars, organic acids, and volatile compounds can affect the chemical and sensory characteristics of the plants, such as microbial stability, pH, and sweetness. Monosaccharides such as rhamnose, fucose, mannose, fructose, arabinose, galactose, xylose, glucose, among others, have been reported in *Annona squamosa* species (Dare et al., 2021), whereas in *Annona muricata* and *Annona cherimola* only sucrose, fructose, and glucose were identified (Chimbevo & Essuman, 2019; Guevara et al., 2019).

Fatty acids are important sources of energy, being also essential components of cell membranes (Ibarguren et al., 2014). Some unsaturated fatty acids, such as linoleic and alpha-linolenic acids, are considered essential because the human organism cannot produce them and must obtain them from the diet. Again, very few studies report the occurrence of these compounds in species of the Annonaceae family.

In *Annona cherimola*, *Annona muricata*, and *Annona squamosa*, for example, oleic acid, a monounsaturated fatty acid linked to lower cholesterol levels and a reduced risk of heart disease, was identified (Luzia & Jorge, 2012). Another fatty acid frequently found in

**Table 4**  
Vitamin contents (mg/100 g dw) of African Annonaceae species.

Scientific name	Plant Parts	Vitamin C	Vitamin E	Thiamin	Riboflavin	Niacin	Cabalamine	Reference
<i>Annona cherimola</i> Mill.	Whole fruit	12.6–25.4	0.27	0.09–0.11	0.11–0.13	0.65–1.0	0.12	(Fuentes et al., 2022)
<i>Annona muricata</i> L.	Whole fruit	1.98 ± 0.01	–	0.11	0.05	1.28	–	(Badrie & Schauss, 2010)
<i>Annona squamosa</i> L.	Fruit pulp	25.6–58.8	–	0.10	0.06	0.89	–	(Fuentes et al., 2022)
<i>Monodora myristica</i> (Gaertn.) Dunal	Seeds	9.9 ± 1.7	5.5 ± 1.4	–	–	–	–	(Nkwocha et al., 2019)
<i>Xylopia aethiopica</i> (Dunal) A.Rich.	Whole fruit	1.83 ± 0.03	0.62 ± 0.02	–	–	–	–	(Fategbe et al., 2021)



Annonaceae species is linoleic acid, a polyunsaturated fatty found in species such as *Annona muricata* and *Annona squamosa*, this being considered an important compound in cardiovascular health, in addition to being involved in the inflammatory process and immunity (Rana, 2015). Other fatty acids, such as margaric, eicosanoic, palmitic, heneicosanoic, and stearic acids, among others, were reported for *Annona muricata* and *Annona squamosa* seeds (Luzia & Jorge, 2012). In these species, (Guevara et al., 2019) were able to identify considerable levels of lactic, acetic, citric, and oxalic acids. Also, according to Elagbar et al. (2016), the main monounsaturated and saturated acids found in *Annona muricata* seeds were oleic (39.2%) and palmitic acids (19.1–19.2%), respectively, and  $\alpha$ -linolenic acid (1.2%) as a polyunsaturated fatty acid.

Vitamin E, in turn, is a fat-soluble organic compound (Galli et al., 2022), that behaves as a natural antioxidant found in several types of foods (Yanishlieva & Marinova, 2001). They are important for human health as they help to protect human cells against oxidative damage caused by free radicals (Devasagayam et al., 2004). Recent studies have shown that some Annonaceae species, including *Annona cherimola*, *Monodora myristica*, and *Xylopia aethiopica* also contain different concentrations of Vitamin E, with *M. myristica* standing out as the one with highest concentrations (5.5 mg/100 g dw) (Nkwocha et al., 2019).

## 5. Identification of edible and non-edible Annonaceae species: Toxicology problems

All over the world, plants are a source of food for many living organisms, including humans, being the basis of the diet over several centuries. Moreover, it is estimated that 80% of the world's population lives in developing countries and uses herbal medicines for basic health care. Some of these species have also been used in developed countries, including in Europe (Braun et al., 2010; Ekor, 2014). But despite the nutritional and medicinal potential of such plants, many of them are poisonous or even deadly when ingested (Wertz & Wynn, 2014).

Particularly, in rural areas, it must be considered that most of these plants have not been exhaustively examined to guarantee their safe use and to prevent possible adverse side effects associated (Kristanc & Kreft, 2016). Several studies have explored the pharmacological properties of the Annonaceae family, although many toxicologic aspects have been ignored, being the incorrect identification of plants for food, the main cause of poisoning (Cornara et al., 2018; Mezzasalma et al., 2017).

Although assessing the bioactivity of plant compounds is the basis for their therapeutic use, it is also important to consider the results of toxicological tests, and to evaluate whether the potentially harmful effects of products derived from these plants override the expected benefits (Gavamukulya et al., 2017). In the past, only morphological characteristics were used, to visually identify the species. Currently, several methods that encompass DNA and other nucleic acids analyses are employed (Miller et al., 2016).

Regarding toxicity, some studies have shown a positive correlation between the consumption of *Annona muricata* fruits and the occurrence of Parkinson's disease (Shaw & Höglinger, 2008). Likewise, toxicity studies carried out by Tan et al. (2007) with an aqueous extract of the stem bark of *Annickia chlorantha* administered orally, revealed histopathological alterations in Swiss albino mice, indicating that medium- and long-term use at doses >500 mg/kg can eventually cause lung, liver, and kidney problems. Some authors have been attributed the toxicity of Annonaceae family to the presence of acetogenins, one of the major classes of bioactive compounds in some species, which possess unique structures and powerful cytotoxic properties, with potential applications as insecticides, antiparasitic, and as a highly interesting new generation of antitumor drugs (García-Aguirre et al., 2008; Hernández-Fuentes et al., 2019). Therefore, it is crucial to understand the mechanisms of action of acetogenins, not only to explore their therapeutic potential but also to assess and mitigate the risks associated with their toxicity.

The mechanism of action of acetogenins involves several steps: i) Inhibition of Mitochondrial Complex I: Acetogenins have been found to

selectively inhibit complex I of the electron transport chain in mitochondria. This inhibition disrupts the flow of electrons, leading to a decrease in ATP (adenosine triphosphate) production and ultimately resulting in cellular dysfunction and apoptosis (programmed cell death). By targeting mitochondrial function, acetogenins induce cell death specifically in cancer cells, which rely heavily on mitochondrial respiration for their energy needs (Grba et al., 2022). ii) Disruption of ATP Production: By inhibiting mitochondrial complex I, acetogenins interfere with the process of oxidative phosphorylation, which is crucial for the generation of ATP, the energy currency of the cell. Deprived of ATP, cancer cells are unable to sustain their metabolic needs and undergo apoptosis (Grba et al., 2022). iii) Induction of Apoptosis: Acetogenins trigger apoptotic pathways in cancer cells. Apoptosis is a tightly regulated process of programmed cell death essential for maintaining tissue homeostasis. Acetogenins induce apoptosis through various mechanisms, including activation of caspases (enzymes that orchestrate cell death), disruption of mitochondrial membrane integrity, and modulation of pro-apoptotic and anti-apoptotic proteins (Han et al., 2015). iv) Selective Toxicity to Cancer Cells: One of the remarkable characteristics of acetogenins is their selective toxicity towards cancer cells while sparing normal cells. This selectivity is attributed to the higher energy demands and increased susceptibility of cancer cells to mitochondrial dysfunction compared to normal cells (Jacobo-Herrera et al., 2019).

Overall, the mechanism of action of acetogenins describe above, making them promising candidates for the development of anticancer therapeutics. However, further research is needed to fully elucidate their therapeutic potential and safety profile.

## 6. Conclusion and future perspectives

In Africa, a large part of the population still depends on medicinal plants for basic health care due to their availability, accessibility, and effectiveness in local communities. This review gathers valuable information about species of the Annonaceae family occurring in Africa that are used as food and in traditional medicine, as well as in other sectors such as the timber industry. The presented data clearly shows that this family encompasses a wide variety of edible and non-edible plant species, many of which are extremely relevant at the medicinal and nutritional levels. The data presented in Table S1, show the wide range of therapeutic possibilities offered by different genera and species of the Annonaceae family through different manipulation and administration methods. This is an excellent starting point for in-depth scientific studies of poorly known plants with high therapeutic potential and social importance in communities with little access to health care. Species such as *Annona muricata*, *Annona cherimola*, *Annona mucosa*, and *Annona reticulata* provide edible fruits, and oils can be extracted from the seeds, leaves, and fruits of species such as *Greenwayodendron suaveolens*, *Annona senegalensis*, *Annona squamosa*, *Monodora tenuifolia*, *Uvaria chamae* and *Xylopia staudtii*. On the other hand, wood from other species, such as *Xylopia acutiflora*, *Xylopia rubescens*, *Xylopia staudtii*, *Duguetia staudtii*, *Uvariopsis dioica* and *Uvariopsis korupensis* are used by local communities in the construction of houses, huts, canoes, and tools for domestic purposes.

Phytochemical studies, in turn, have reported the isolation of different bioactive compounds from the seeds, leaves, bark, and pulp of Annonaceae species, such as acetogenins, flavonoids, and alkaloids. Other species are also important sources of essential oils with high antimicrobial and insecticidal activities, used as traditional medicines to treat malaria, diabetes, rheumatism, and fever, among several other illnesses, and as insecticides.

The different parts of some species are rich in nutrients, antioxidants, and other bioactive compounds, which makes them potential sources for the development of new bio-based drugs, extremely relevant for the pharmaceutical industry, and functional foods. Furthermore, these medicinal and nutritional characteristics are of paramount importance in developing countries, as in Africa, where access to food and medicines is

limited and people must use the natural resources locally available to meet their needs. Some of the compounds isolated from these plants have shown promising properties in *in vitro* and *in vivo* assays, including anticancer, anti-malarial, anti-parasitic, anti-inflammatory, and neuro-protective activities, among several other therapeutic effects. Although many Annonaceae species have a huge potential as sources of bioactive compounds, it is important to consider that some of them may contain harmful compounds and thus be toxic if consumed in large quantities.

Among the countries of sub-Saharan Africa, Cameroon and Nigeria are the ones that have conducted the most studies on Annonaceae species, encompassing traditional uses and phytochemical properties. However, there is still a significant dearth of research on the nutritional and chemical profiles of many species. Therefore, it is necessary to conduct further studies on species within this family, considering their degree of importance in African communities and beyond. Throughout this study, we found that the most exploited species in Africa are *Annickia chlorantha*, *Annona muricata*, *Annona senegalensis*, *Annona squamosa*, *Duguetia staudtii*, *Monodora myristica*, *Uvaria chamae*, *Xylopia aethiopica*, *Xylopia longipetala* and *Xylopia staudtii*.

Therefore, future research on Annonaceae species should be focused on the development of sustainable cultivation practices, preservation of their natural habitats, identification of further safe and potentially beneficial species, and rational exploitation of their bioactive compounds for nutritional and pharmaceutical purposes. Therefore, encouraging the study of African Annonaceae species can boost the discovery of new bioactive compounds with therapeutic potential, new sources of micro- and macronutrients, and expand the range of food products and phytochemicals available to the growing and more demanding world population.

#### CRediT authorship contribution statement

**Josefa Rangel:** Writing – original draft, Investigation. **Sílvia Catarina:** Writing – review & editing, Investigation. **José Carlos Costa:** Writing – review & editing, Supervision. **Maria M. Romeiras:** Writing – review & editing, Visualization, Supervision, Conceptualization. **Ângela Fernandes:** Writing – original draft, Investigation.

#### Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### Data availability

No data was used for the research described in the article.

#### Acknowledgments

The authors are grateful to the Foundation for Science and Technology (FCT, Portugal) for financial support from the FCT/MCTES (PIDDAC) national funds to CIMO (UIDB/00690/2020 (DOI: [10.54499/UIDB/00690/2020](https://doi.org/10.54499/UIDB/00690/2020)) and UIDP/00690/2020 (DOI: [10.54499/UIDP/00690/2020](https://doi.org/10.54499/UIDP/00690/2020))), SusTEC (LA/P/0007/2020 (DOI: [10.54499/LA/P/0007/2020](https://doi.org/10.54499/LA/P/0007/2020))), LEAF (UIDB/04129/2020 and UIDP/04129/2020) and cE3c (UIDB/00329/2020 (DOI: [10.54499/UIDB/00329/2020](https://doi.org/10.54499/UIDB/00329/2020))) and for the national funding by FCT and P.I. in the form of the institutional scientific employment program for the contracts of Â. Fernandes (DOI: [10.54499/CEECINST/00016/2018/CP1505/CT0008](https://doi.org/10.54499/CEECINST/00016/2018/CP1505/CT0008)), and the PhD fellowships PRT/BD/152088/2021 and 2021.04585.BD of J. Rangel and Â. Liberal, respectively. S. Catarino was funded by GenoCash Project (PTDC/ASP-AGR/0760/2020).

#### Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.foodchem.2024.139048>.

#### References

- Abdualrahman, M. A. Y., Ma, H., Zhou, C., Ahmed Yagoub, A. E. G., Ali, A. O., Tahir, H. E., & Wali, A. (2019). Postharvest physicochemical properties of the pulp and seed oil from *Annona squamosa* L. (Gishta) fruit grown in Darfur region, Sudan. *Arabian Journal of Chemistry*, 12, 4514–4521. <https://doi.org/10.1016/j.arabjc.2016.07.008>
- Adesawo, J. K., Akinloye, A. A., Otemuyiwa, I. O., & Akinpelu, D. A. (2020). Chemical characteristics and biological activities of *Annona squamosa* fruit pod and seed extracts. *Journal of Exploratory Research in Pharmacology*, 6, 5–15. <https://doi.org/10.14218/JERP.2020.00019>
- Adesokan, A. A., & Akanji, M. A. (2010). Antimalarial bioactivity of *Enantia chlorantha* stem bark. *Pharmacology and Therapeutics*, 1, 441–447.
- Aggarwal, V., Joshi, N., & Varghese, J. (2018). Study of bioactive compounds from Methanolic whole fruit extracts of *Annona reticulata* and *Annona squamosa* by GC-MS and evaluation of antioxidant activity. *Innovations in Food, Environment and Healthcare (NCIFEH-2018)*, 85.
- Agiriga, A., & Siwela, M. (2017). *Monodora myristica* (Gaertn.) Dunal: A plant with multiple food, health and medicinal applications: A review. *American Journal of Food Technology*, 12, 271–284. <https://doi.org/10.3923/ajft.2017.271.284>
- Aiyelaja, A. A., & Bello, O. A. (2006). Ethnobotanical potentials of common herbs in Nigeria: A case study of Enugu state. *Educational Research Review*, 1, 16–22.
- Akendengue, B., Ngou-Milama, E., Bouroubou-Bouroubou, H., Essouma, J., Roblot, F., Gleye, C., Laurens, A., Hocquemiller, R., Loiseau, P., & Bories, C. (2003). Acaricidal activity of *Uvaria versicolor* and *Uvaria klaineana* (Annonaceae). *Phytotherapy Research*, 17, 364–367. <https://doi.org/10.1002/ptr.1158>
- Akomolafe, S. F., & Ajayi, O. B. (2015). A comparative study on antioxidant properties, proximate and mineral compositions of the peel and pulp of ripe *Annona muricata* (L.) fruit. *International Food Research Journal*, 22, 2381–2388.
- Al-Nemari, R., Bacha, A. B., Al-Senaidey, A., Almutairi, M. H., Arafah, M., Al-Saran, H., ... Semlali, A. (2022). Cytotoxic effects of *Annona squamosa* leaves against breast cancer cells via apoptotic signaling proteins. *Journal of King Saud University, Science*, 34, Article 102013. <https://doi.org/10.1016/j.jksus.2022.1>
- Arena, M. E., Povelonis, I. S., Borroni, V., Constenla, D., & Radice, S. (2021). Changes in physicochemical properties at different development stages of *Hexachlamys edulis* fruit, an underutilized South American species. *Heliyon*, 7. <https://doi.org/10.1016/j.heliyon.2021.e08323>, 1–11.
- Atasie, V. N., Akinhanmi, T. F., & Ojiodu, C. C. (2009). Proximate analysis and physicochemical properties of groundnut (*Arachis hypogaea* L.). *Pakistan Journal of Nutrition*, 8, 194–197. <https://doi.org/10.3923/pjn.2009.194.197>
- Ayessou, N. C., Gueye, M., Dioh, E., Konteye, M., Cissé, M., & Dornier, M. (2009). Nutritive composition and energy contribution of the fruit of *Maerua pseudopetalosa*, a food extender in Senegal. *Fruits*, 64, 147–156. <https://doi.org/10.1051/fruits/2009010>
- Badrie, N., & Schauss, A. G. (2010). Soursop (*Annona muricata* L.): Composition, nutritional value, medicinal uses, and toxicology in *Bioactive Foods in Promoting Health*. Chapter, 39, 621–643. <https://doi.org/10.1016/B978-0-12-374628-3.00039-6>
- Ben-Othman, S., Joudou, I., & Bhat, R. (2020). Bioactives from Agri-food wastes: Present insights and future challenges. *Molecules*, 25, 510. <https://doi.org/10.3390/molecules25030510>
- Bhardwaj, R., Pareek, S., Sagar, N. A., & Vyas, N. (2019). Bioactive compounds of *Annona* BT - bioactive compounds in underutilized fruits and nuts. In H. N. Murthy, & V. A. Bapat (Eds.), *Reference Series in Phytochemistry* (pp. 1–26). [https://doi.org/10.1007/978-3-030-06120-3\\_5-1](https://doi.org/10.1007/978-3-030-06120-3_5-1)
- Biba, V. S., Amily, A., Sangeetha, S., & Remani, P. (2014). Anticancer, antioxidant and antimicrobial activity of Annonaceae family. *World Journal of Pharmacy and Pharmaceutical Sciences*, 3, 1595–1604.
- Borquaye, L. S., Darko, G., Laryea, M. K., Gasu, E. N., Amponsah, N. A. A., & Appiah, E. N. (2017). Nutritional and anti-nutrient profiles of some Ghanaian spices. *Cogent Food & Agriculture*, 3, 1348185. <https://doi.org/10.1080/23311932.2017.1348185>
- Braun, L. A., Tiralongo, E., Wilkinson, J. M., Spitzer, O., Bailey, M., Poole, S., & Dooley, M. (2010). Perceptions, use and attitudes of pharmacy customers on complementary medicines and pharmacy practice. *BMC Complementary and Alternative Medicine*, 10, 1–7. <https://doi.org/10.1186/1472-6882-10-38>
- Catarino, L., Havik, P. J., & Romeiras, M. M. (2016). Medicinal plants of Guinea-Bissau: Therapeutic applications, ethnic diversity and knowledge transfer. *Journal of Ethnopharmacology*, 183, 71–94. <https://doi.org/10.1016/j.jep.2016.02.032>
- Catarino, S., Rangel, J., Darbyshire, I., Costa, E., Duarte, M. C., & Romeiras, M. M. (2021). Conservation priorities for African *Vigna* species: Unveiling Angola's diversity hotspots. *Global Ecology and Conservation*, 25, 1415. <https://doi.org/10.1016/j.gecco.2020.e01415>
- Chimbevo, L. M., & Essuman, S. (2019). Preliminary screening of nutraceutical potential of fruit pulp, peel and seeds from *Annona squamosa* (L.) and *Annona muricata* (L.) growing in coast region of Kenya. *American Journal of BioScience*, 7, 58–70. <https://doi.org/10.11648/j.ajbio.20190703.11>

- Chukwuma, E. R., & Uzoma, N. O. (2013). Biochemical studies on Nigerian *Monodora tenuifolia* seed. *American Journal of Agricultural and Biological Sciences*, 8, 257. <https://doi.org/10.3844/ajabssp.2013.257.267>
- Cordeiro, N., Sousa, L., Freitas, N., & Gouveia, M. (2013). Changes in the mesocarp of *Annona cherimola* mill. "Madeira" during postharvest ripening. *Postharvest Biology and Technology*, 85, 179–184. <https://doi.org/10.1016/j.postharvbio.2013.05.014>
- Coria-Téllez, A. V., Montalvo-González, E., Yahia, E. M., & Obledo-Vázquez, E. N. (2018). *Annona muricata*: A comprehensive review on its traditional medicinal uses, phytochemicals, pharmacological activities, mechanisms of action and toxicity. *Arabian Journal of Chemistry*, 11, 662–691. <https://doi.org/10.1016/j.arabj.2016.01.004>
- Cornara, L., Smeriglio, A., Frigerio, J., Labra, M., Di Gristina, E., Denaro, M., Mora, E., & Trombetta, D. (2018). The problem of misidentification between edible and poisonous wild plants: Reports from the Mediterranean area. *Food and Chemical Toxicology*, 119, 112–121. <https://doi.org/10.1016/j.fct.2018.04.066>
- Costa, E., & Pedro, M. (2013). *Plantas medicinais de Angola*. Luanda: Centro de Botânica da Faculdade de Ciências da Universidade Agostinho Neto.
- Costa, E. V., Soares, L. D. N., Chaar, J. D. S., Silva, V. R., Santos, L. D. S., Koolen, H. H. F., ... Bezerra, D. P. (2021). Benzylated dihydroflavones and isoquinoline-derived alkaloids from the bark of *Diclinanona calycina* (Annonaceae) and their cytotoxicities. *Molecules*, 26, 3714. <https://doi.org/10.3390/molecules26123714>
- Couvreux, T. L. (2008). *Revealing the secrets of African Annonaceae: systematics, evolution and biogeography of the syncarpous genera Isolona and Monodora* (p. 307). The Netherlands: Wageningen University and Research, PhD thesis Wageningen University.
- Couvreux, T. L. P., Gereau, R. E., Wieringa, J. J., & Richardson, J. E. (2006). Description of four new species of *Monodora* and *Isolona* (Annonaceae) from Tanzania and an overview of Tanzanian Annonaceae diversity. *Adansonia*, 28, 243–266.
- Daniels, A. O., & Ibiyemi, D. A. (2021). Identification of the functional groups, radical scavenging ability and antimicrobial properties of the extract of *Cleistopholis patens*. *Achievers Journal of Scientific Research*, 3, 105–115.
- Dare, C., Oyedapo, O., Akinlalu, A., Komolafe, I., Fajobi, A., & Ogunsusi, M. (2021). Genotoxic activities of polysaccharides from Cotyledon and coat of fermented and unfermented *Annona squamosa* L. Seed. *Egyptian Academic Journal of Biological Sciences, H. Botany*, 12, 189–207. <https://doi.org/10.21608/eajbsh.2021.199654>
- Dawodu, A. O., Moses, U. D., Apena, A., Adetoro, A., & Dairo, J. O. (2014). The proximate evaluation and Phytochemistry of *Enantia chlorantha* stem bark in aqueous and Ethanolic extract. *Middle-East Journal of Scientific Research*, 21, 2145–2148. <https://doi.org/10.5829/idosi.mejsr.2014.21.11.21842>
- Devasagayam, T. P. A., Tilak, J. C., Boloor, K. K., Sane, K. S., Ghaskadbi, S. S., & Lele, R. D. (2004). Free radicals and antioxidants in human health: Current status and future prospects. *Journal of the Association of Physicians of India*, 52, 4.
- Didier, A. J., Stiene, J., Fang, L., Watkins, D., Dworkin, L. D., & Creeden, J. F. (2023). Antioxidant and anti-tumor effects of dietary vitamins A, C, and E. *Antioxidants*, 12, 1–26. <https://doi.org/10.3390/antiox12030632>
- Ehara, H., Toyoda, Y., & Johnson, D. V. (2018). Sago palm: Multiple contributions to food security and sustainable livelihoods. In *I. Sago palm: Multiple contributions to food security and sustainable livelihoods*. Springer Nature (pp. 1–317). Japan: Hiroshi Ehara Applied Social System Institute of Asia, International Cooperation Center for Agricultural Education Nagoya University Nagoya. <https://doi.org/10.1007/978-981-10-269-9>
- Ekor, M. (2014). The growing use of herbal medicines: Issues relating to adverse reactions and challenges in monitoring safety. *Frontiers in Pharmacology*, 177, 1–10. <https://doi.org/10.3389/fphar.2013.00177>. Abidemi James Akindele, University of Lagos, Nigeria. 4.
- Elagbar, Z. A., Naik, R. R., Shakya, A. K., & Bardaweel, S. K. (2016). Fatty acids analysis, antioxidant and biological activity of fixed oil of *Annona muricata* L. Seeds. *Journal of Chemistry*, 2016, 1–6. <https://doi.org/10.1155/2016/6948098>
- El-Ramady, H., Hajdú, P., Törös, G., Badgar, K., Ilanaj, X., Kiss, A., ... Prokisch, J. (2022). Plant nutrition for human health: A pictorial review on plant bioactive compounds for sustainable agriculture. *Sustainability*, 14, 8329. <https://doi.org/10.3390/su14148329>
- Erkens, R. H., Blanpain, L. M., Carrascosa Jara, I., Runge, K., Verspagen, N., Cosiaux, A., & Couvreur, T. L. (2023). Spatial distribution of Annonaceae across biomes and anthromes: Knowledge gaps in spatial and ecological data. *Plants, People, Planet*, 5, 520–535. <https://doi.org/10.1002/ppp3.10321>
- Fall, D., Pimentel, L., Champy, P., Gleye, C., Laurens, A., & Hocquemiller, R. (2006). A new adjacent bis-tetrahydrofuran annonaceous acetogenin from the seeds of *Uvaria chamae*. *Planta Medica*, 72, 938–940. <https://doi.org/10.1055/s-2006-941542>
- Fategbe, M. A., Awvioroko, O. J., & Ibukun, E. O. (2021). Comparative biochemical evaluation of the proximate, mineral, and phytochemical constituents of *Xylopia aethiopica* whole fruit, seed, and pericarp. *Preventive Nutrition and Food Science*, 26, 219. <https://doi.org/10.3746/pnf.2021.26.2.219>
- Floegel, A., Kim, D. O., Chung, S. J., Koo, S. I., & Chun, O. K. (2011). Comparison of ABTS/DPPH assays to measure antioxidant capacity in popular antioxidant-rich US foods. *Journal of Food Composition and Analysis*, 24, 1043–1048. <https://doi.org/10.1016/j.jfca.2011.01.008>
- Fournier, G., Leboeuf, M., & Cavé, A. (1999). Annonaceae essential oils: A review. *Journal of Essential Oil Research*, 11, 131–141. <https://doi.org/10.1080/10412905.1999.9701092>
- Fuentes, L. M., González, E. M., Magaña, M. d. L. G., Esparza, L. M. A., González, Y. N., Villagrán, Z., ... Flores, D. A. M. (2022). Current situation and perspectives of fruit Annonaceae in Mexico: Biological and agronomic importance and bioactive properties. *Plants*, 11, 7. <https://doi.org/10.3390/plants11010007>
- Galli, F., Bonomini, M., Bartolini, D., Zatini, L., Reboldi, G., Marcantonini, G., ... Di Pietro, N. (2022). Vitamin E (alpha-tocopherol) metabolism and nutrition in chronic kidney disease. *Antioxidants*, 11, 989. <https://doi.org/10.3390/antiox11050989>
- García-Aguirre, K. K., Zepeda-Vallejo, L. G., Ramón-Gallegos, E., Álvarez-González, & Madrigal-Bujaidar, E. (2008). Genotoxic and cytotoxic effects produced by acetogenins obtained from *Annona cherimolia* Mill. *Biological and Pharmaceutical Bulletin*, 31, 2346–2349. <https://doi.org/10.1248/bpb.31.2346>
- Gavamukulya, Y., Wamunyokoli, F., & El-Shemy, H. A. (2017). *Annona muricata*: Is the natural therapy to most disease conditions including cancer growing in our backyard? A systematic review of its research history and future prospects. *Asian Pacific Journal of Tropical Medicine*, 10, 835–848. <https://doi.org/10.1016/j.apjtm.2017.08.009>
- George, V. C., Kumar, D. R., Rajkumar, V., Suresh, P. K., & Kumar, R. A. (2012). Quantitative assessment of the relative antineoplastic potential of the n-butanolic leaf extract of *Annona muricata* Linn. In normal and immortalized human cell lines. *Asian Pacific Journal of Cancer Prevention*, 13, 699–704. <https://doi.org/10.7314/apjcp.2012.13.2.699>
- Goncharova, N. A., & Merzlyakova, N. V. (2022). Food shortages and hunger as a global problem. *Food Science And Technology Brazil*, 42, 70621. <https://doi.org/10.1590/fst.70621>
- González-Aguero, M., Pardo, L. T., Zamudio, M. S., Contreras, C., Undurraga, P., & Defilippi, B. G. (2016). The unusual acid-accumulating behavior during ripening of cherimoya (*Annona cherimola* mill.) is linked to changes in transcription and enzyme activity related to citric and malic acid metabolism. *Molecules*, 21, 398. <https://doi.org/10.3390/molecules21050398>
- Grba, D. N., Blaza, J. N., Bridges, H. R., Agip, A.-N. A., Yin, Z., Murai, M., ... Hirst, J. (2022). Cryo-electron microscopy reveals how acetogenins inhibit mitochondrial respiratory complex I. *Journal of Biological Chemistry*, 298, Article 101602. <https://doi.org/10.1016/j.jbc.2022.101602>
- Guevara, M., Tejera, E., Granda-Albuja, M. G., Iturralde, G., Chisaguano-Tonato, M., Granda-Albuja, S., ... Alvarez-Suarez, J. M. (2019). Chemical composition and antioxidant activity of the main fruits consumed in the western coastal region of Ecuador as a source of health-promoting compounds. *Antioxidants*, 8, 387. <https://doi.org/10.3390/antiox8090387>
- Han, B., Wang, T.-D., Shen, S.-M., Yu, Y., Mao, C., Yao, Z.-J., & Wan, L.-S. (2015). Annonaceous acetogenin mimic AA005 induces cancer cell death via apoptosis inducing factor through a caspase-3-independent mechanism. *BMC Cancer*, 15, 139. <https://doi.org/10.1186/s12885-015-1133-0>
- Handayani, T. (2018). Diversity, potential and conservation of Annonaceae in Bogor botanic gardens, Indonesia. *Biodiversitas*, 19, 541–553. <https://doi.org/10.13057/biodiv/d190230>
- Handayani, T., & Yuzammi. (2021). The family of Annonaceae: The important role in forest ecosystems and human being life. *IOP Conference Series: Earth and Environmental Science*, 914, 12062. <https://doi.org/10.1088/1755-1315/914/1/012062>
- Hernández-Fuentes, G. A., García-Argáez, A. N., Campos, A. L. P., Delgado-Enciso, I., Muñoz-Valencia, R., Martínez-Martínez, F. J., ... Parra-Delgado, H. (2019). Cytotoxic acetogenins from the roots of *Annona purpurea*. *International Journal of molecular Sciences*, 20, 1870. <https://doi.org/10.3390/ijms20081870>
- Ibarguren, M., López, D. J., & Escribá, P. V. (2014). The effect of natural and synthetic fatty acids on membrane structure, microdomain organization, cellular functions and human health. *Biochimica et Biophysica Acta, Biomembranes*, 1838, 6. <https://doi.org/10.1016/j.bbame.2013.12.021>
- Imo, C., Yakubu, O., Imo, N., Udegbunwa, I., & Onukwugha, O. (2018). Chemical composition of *Xylopia aethiopica* fruits. *American Journal of Physiology, Biochemistry and Pharmacology*, 7, 48–53. <https://doi.org/10.5455/ajpbp.20180521064020>
- Ironi, E. A., Aroyehun, T. M., Anyiam, A. F., & Lal, M. K. (2023). Phenolics profile, anti-nephrolithiasis, and antioxidant activities of *Monodora myristica* seed: Impact of endogenous proteins and lipids. *Food Production, Processing and Nutrition*, 5, 52. <https://doi.org/10.1186/s43014-023-00167-8>
- Ita, B. (2017). Antioxidant activity of *Cnestis ferruginea* and *Uvaria chamae* seed extracts. *British Journal of Pharmaceutical Research*, 16, 1–8. <https://doi.org/10.9734/bjpr/2017/32924>
- Jacobo-Herrera, N., Pérez-Plasencia, C., Castro-Torres, V. A., Martínez-Vázquez, M., González-Esquina, A. R., & Zentella-Dehesa, A. (2019). Selective Acetogenins and their potential as anticancer agents. *Frontiers in Pharmacology*, 10, 783. <https://doi.org/10.3389/fphar.2019.00783>
- Jalil, J., Attiq, A., Hui, C. C., Yao, L. J., & Zakaria, N. A. (2020). Modulation of inflammatory pathways, medicinal uses and toxicities of *Uvaria* species: Potential role in the prevention and treatment of inflammation. *Inflammopharmacology*, 28, 1195–1218. <https://doi.org/10.1007/s10787-020-00734-2>
- Jiménez, V. M., Gruschwitz, M., Schweiggert, R. M., Carle, R., & Esquivel, P. (2014). Identification of phenolic compounds in soursop (*Annona muricata*) pulp by high-performance liquid chromatography with diode array and electrospray ionization mass spectrometric detection. *Food Research International*, 65, 42–46. <https://doi.org/10.1016/j.foodres.2014.05.051>
- Kasali, F. M., Kadima, J. N., Peter, E. L., Mtewa, A. G., Ajayi, C. O., Tusiimire, J., ... Agaba, A. G. (2021). Antidiabetic medicinal plants used in Democratic Republic of Congo: A critical review of ethnopharmacology and bioactivity data. *Frontiers in Pharmacology*, 12, Article 757090. <https://doi.org/10.3389/fphar.2021.757090>
- Kayser, O. (2018). Ethnobotany and medicinal plant biotechnology: From tradition to modern aspects of drug development. *Planta Medica*, 84, 834–838. <https://doi.org/10.1055/a-0631-3876>
- Keinan, E., & Sinha, S. C. (2002). Oxidative polycyclizations with rhenium (VII) oxides. *Pure and Applied Chemistry*, 74, 93–105. <https://doi.org/10.1351/pac200274010093>



- Kn, C., Shaji, C., & Thomas, B. (2016). Evaluation of major phytochemical constituents of two edible fruit yielding species of Annonaceae: *Annona muricata* L. and *Annona reticulata* L. *Journal of Medicinal Plants Studies*, 4, 198–202.
- Koudouvo, K., Karou, D. S., Kokou, K., Essien, K., Aklikokou, K., Glitho, I. A., ... Gbeassor, M. (2011). An ethnobotanical study of antimalarial plants in Togo maritime region. *Journal of Ethnopharmacology*, 134, 183–190. <https://doi.org/10.1016/j.jep.2010.12.011>
- Kristanc, L., & Kreft, S. (2016). European medicinal and edible plants associated with subacute and chronic toxicity part I: Plants with carcinogenic, teratogenic and endocrine-disrupting effects. *Food and Chemical Toxicology*, 92, 150–164. <https://doi.org/10.1016/j.fct.2016.04.007>
- Lamperti, A. M., French, A. R., Dierenfeld, E. S., Fogiel, M. K., Whitney, K. D., Stauffer, D. J., ... Parker, V. T. (2014). Diet selection is related to breeding status in two frugivorous hornbill species of Central Africa. *Journal of Tropical Ecology*, 30, 273–290. <https://doi.org/10.1017/S0266467414000236>
- Lee, C. H., Lee, T. H., Ong, P. Y., Wong, S. L., Hamdan, N., Elgharabawy, A. A. M., & Azmi, N. A. (2021). Integrated ultrasound-mechanical stirrer technique for extraction of total alkaloid content from *Annona muricata*. *Process Biochemistry*, 109, 104–116. <https://doi.org/10.1016/j.procbio.2021.07.006>
- Liaw, C. C., Wu, T. Y., Chang, F. R., & Wu, Y. C. (2010). Historic perspectives on Annonaceous acetogenins from the chemical bench to preclinical trials. *Planta Medica*, 76, 1390–1404. <https://doi.org/10.1055/s-0030-1250006>
- López-Martínez, C. R., García Mateos, M. D. R., & Martínez Damían, M. T. (2022). Nutritional and nutraceutical quality of the fruit of three species of Annonaceae: Soursop, cherimoya and chincuya. *Nova Scientia*, 14, 1–15. <https://doi.org/10.21640/ns.v14i28.2925>
- Luzia, D. M. M., & Jorge, N. (2012). Soursop (*Annona muricata* L.) and sugar apple (*Annona squamosa* L.): Antioxidant activity, fatty acids profile and determination of tocopherols. *Nutrition & Food Science*, 42, 434–441. <https://doi.org/10.1108/00346651211277690>
- Mabhaudhi, T., Chimonyo, V. G. P., & Modi, A. T. (2017). Status of underutilised crops in South Africa: Opportunities for developing research capacity. *Sustainability*, 9, 1569. <https://doi.org/10.3390/su9091569>
- Maeda, G., Gillissen, P. J., Bourgard, C., Wal, J. V. D., Munissi, J. J. E., Nyandoro, S. S., & Erdélyi, M. (2022). Polyoxxygenated cyclohexene derivatives and flavonoids from the leaves of *Uvaria pandensis*. *Fitoterapia*, 158, 105170.
- Maia, D. S., Lopes, C. F., Saldanha, A. A., Silva, N. L., Sartori, Â. L. B., Carollo, C. A., ... de Siqueira, J. M. (2020). Larvicidal effect from different Annonaceae species on *Culex quinquefasciatus*. *Environmental Science and Pollution Research*, 27, 36983–36993. <https://doi.org/10.1007/s11356-020-08997-6>
- Makhuvele, R., Foubert, K., Hermans, N., Pieters, L., Verschaeve, L., & Elgorashi, E. (2022). Protective effects of methanolic leaf extracts of *Monanthotaxis caffra* against aflatoxin B1-induced hepatotoxicity in rats. *Onderstepoort Journal of Veterinary Research*, 89, 1968. <https://doi.org/10.4102/ojvr.v89i1.1968>
- Malan, D. F., Neuba, D. F. R., & Kouakou, K. L. (2015). Medicinal plants and traditional healing practices in ehophile people, around the aby lagoon (eastern littoral of Côte d'Ivoire). *Journal of Ethnobiology and Ethnomedicine*, 11, 1–18. <https://doi.org/10.1186/s13002-015-0004-8>
- Mannino, G., Gentile, C., Porcu, A., Agliassa, C., Caradonna, F., & Berte, C. M. (2020). Chemical profile and biological activity of cherimoya (*Annona cherimola* Mill.) and atemoya (*Annona atemoya*) leaves. *Molecules*, 25, 2612. <https://doi.org/10.3390/molecules25112612>
- Martin, C., Viruel, M. A., Lora, J., & Hormaza, J. I. (2019). Polyploidy in fruit tree crops of the genus *Annona* (Annonaceae). *Frontiers in Plant Science*, 10, 99. <https://doi.org/10.3389/fpls.2019.00099>
- Menezes, E. G. T., Oliveira, E. R., Carvalho, G. R., Guimaraes, I. C., & Queiroz, F. (2019). Assessment of chemical, nutritional and bioactive properties of *Annona crassiflora* and *Annona muricata* wastes. *Food Science and Technology*, 39, 662–672. <https://doi.org/10.1590/fst.22918>
- Mezzasalma, V., Ganopoulos, I., Galimberti, A., Cornara, L., Ferri, E., & Labra, M. (2017). Poisonous or non-poisonous plants? DNA-based tools and applications for accurate identification. *International Journal of Legal Medicine*, 131, 1–19. <https://doi.org/10.1007/s00414-016-1460-y>
- Mfopa, A. N., Mbouna, C. D. J., Tchokouaha, L. R. Y., Tchuente, M. A. T., Kouipou, R. M. T., Fokou, P. V. T., & Boyom, F. F. (2017). *In vitro* and *in vivo* antiparasitic activity of extracts from *Polyalthia suaveolens*, *Uvaria angolensis* and *Monodora tenuifolia* (Annonaceae). *International Journal of Biological and Chemical Sciences*, 11, 118–130. <https://doi.org/10.20944/preprints201612.0044.v1>
- Miller, S. E., Hausmann, A., Hallwachs, W., & Janzen, D. H. (2016). Advancing taxonomy and bioinventories with DNA barcodes. *Philosophical Transactions of the Royal Society, B: Biological Sciences*, 371, 20150339. <https://doi.org/10.1098/rstb.2015.0339>
- Mols, J. B., Gravendeel, B., Chatrou, L. W., Pirie, M. D., Bygrave, P. C., Chase, M. W., & Keßler, P. J. A. (2004). Identifying clades in Asian Annonaceae: Monophyletic genera in the polyphyletic Miliuseae. *American Journal of Botany*, 91, 590–600. <https://doi.org/10.3732/ajb.91.4.590>
- Monon, K., Abdoulaye, T., Karamoko, O., & Adama, C. (2015). Phytochemical composition, antioxidant and antibacterial activities of root of *Uvaria chamae* p. Beauv. (Annonaceae) used in treatment of dysentery in north of Côte d'Ivoire. *International Journal of Pharmacognosy and Phytochemical Research*, 7, 1047–1053.
- Muganza, D. M., Fruth, B., Nzunzu, J. L., Tuentner, E., Foubert, K., Cos, P., ... Pieters, L. (2016). *In vitro* antiprotazoal activity and cytotoxicity of extracts and isolated constituents from *Greenwayodendron suaveolens*. *Journal of Ethnopharmacology*, 193, 510–516. <https://doi.org/10.1016/j.jep.2016.09.051>
- Munissi, J. (2019). Cytotoxic and antimicrobial activities of the constituents of ten Plant species from Tanzania. *Tanzania Journal of Science*, 45, 44–52.
- Nam, J. S., Jang, H. L., & Rhee, Y. H. (2018). Nutritional compositions in roots, twigs, leaves, fruit pulp, and seeds from pawpaw (*Asimina triloba* [L.] Dunal) grown in Korea. *Journal of Applied Botany and Food Quality*, 91, 47–55. <https://doi.org/10.5073/JABFQ.2018.091.007>
- Nguiam, M. P., Wouamba, S. C. N., Longo, F., Kamkumo, R. G., Foweda, L. D. K., Djomeni, P. D. D., ... Dimo, T. (2021). Antibacterial and antishigellosis activity of *Xylopia staudtii* (Engl & Diels), Annonaceae. *Journal of Ethnopharmacology*, 280, Article 114406. <https://doi.org/10.1016/j.jep.2021.114406>
- Nishiyama, Y., Moriyasu, M., Ichimaru, M., Iwasa, K., Kato, A., Mathenge, S. G., ... Juma, F. D. (2004). Quaternary isoquinoline alkaloids from *Xylopia parviflora*. *Phytochemistry*, 65, 939–944. <https://doi.org/10.1016/j.phytochem.2003.12.010>
- Nkwocha, C. C., Okagu, I. U., & Chibugwu, C. C. (2019). Mineral and vitamin contents of *Monodora myristica* (African nutmeg) seeds from Nsukka, Enugu state, Nigeria. *Pakistan Journal of Nutrition*, 18, 308–314. <https://doi.org/10.3923/pjn.2019.308.314>
- Noudogbessi, J. P., Gary-Bobo, M., Adomou, A., Adjalian, E., Alitonou, G. A., Avlessi, F., ... Menut, C. (2014). Comparative chemical study and cytotoxic activity of *Uvariadendron angustifolium* essential oils from Benin. *Natural Product Communications*, 9, 261–264. <https://doi.org/10.1177/1934578X1400900232>
- Nyandoro, S. S. (2017). *In ovo* antiviral activity of the constituents of *Artabotrys monteiroae* and *Artabotrys modestus* against infectious bursal disease and Newcastle disease viruses. *International Journal of Biological and Chemical Sciences*, 11, 3075–3085. <https://doi.org/10.4314/ijbcs.v11i6.41>
- Nyandoro, S. S., Maeda, G., Munissi, J. J. E., Gruhonic, A., Fitzpatrick, P. A., Lindblad, S., ... Erdélyi, M. (2019). A new benzopyranyl cadenane sesquiterpene and other antiparasitic and cytotoxic metabolites from *Cleistochlamys kirkii*. *Molecules*, 24, 2746. <https://doi.org/10.3390/molecules24152746>
- Ofeimun, J. O., Eze, G. I., Okirika, O. M., & Uansejor, S. O. (2013). Evaluation of the hepatoprotective effect of the methanol extract of the root of *Uvaria afzelii* (Annonaceae). *Journal of Applied Pharmaceutical Science*, 3, 125–129. <https://doi.org/10.7324/JAPS.2013.31022>
- Ojokuku, S. A., Apena, A., Dawodu, A. O., & Odunlade, A. K. (2011). Nutritional quality of two medicinal plants (*Anthocleista vogelii* and *Uvaria chamae*) in Western Nigeria. *International Journal of Science & Society Yabatech*, 1, 58–62.
- Okigbo, R. N., & Anyaegbu, C. F. (2021). Underutilized plants of Africa. *Journal of Biology and Nature*, 13, 34.
- Oshomoh, E. O., Obasuyi, E. I., & Eze, F. S. (2022). Proximate composition, phytonutrient analysis and antimicrobial activity of Ethanolic extracts of the young and mature leaves of *Greenwayodendron suaveolens*. *Journal of Applied Sciences and Environmental Management*, 26, 449–452. <https://doi.org/10.4314/jasem.v26i3.11>
- Parmena, D. S., Mgina, C. A., & Joseph, C. C. (2012). Composition of non-volatile oils and antimicrobial activities of extracts from *Monanthotaxis discolor*, and an undescribed *Uvariadendron* species. *Tanzania Journal of Science*, 38, 221–231.
- Pedrali, A., Robustelli Della Cuna, F. S., Grisoli, P., Corti, M., & Brusotti, G. (2019). Chemical composition and antimicrobial activity of the essential oil from the bark of *Xylopia hypolampra*. *Natural Product Communications*, 14, 1–5. <https://doi.org/10.1177/1934578X19857022>
- Pereira, F., Madureira, A. M., Sancha, S., Mulhovo, S., Luo, X., Duarte, A., & Ferreira, M. J. U. (2016). *Cleistochlamys kirkii* chemical constituents: Antibacterial activity and synergistic effects against resistant *Staphylococcus aureus* strains. *Journal of Ethnopharmacology*, 178, 180–187. <https://doi.org/10.1016/j.jep.2015.12.009>
- Perrone, A., Yousefi, S., Salami, A., Papini, A., & Martinelli, F. (2022). Botanical, genetic, phytochemical and pharmaceutical aspects of *Annona cherimola* Mill. *Scientia Horticulturae*, 296, Article 110896. <https://doi.org/10.1016/j.scienta.2022.110896>
- Pimenta, A. C., Rego, S. S., Zuffellato-Ribas, K. C., Nogueira, A. C., & Koehler, H. S. (2013). Morphological characterization of fruits, seeds and seedlings of araticum plant (*Annona crassiflora* Mart - Annonaceae). *Journal of Seed Science*, 35, 524–531. <https://doi.org/10.1590/s2317-15372013000400015>
- Qian, J. Q., Sun, P., Pan, Z. Y., & Fang, Z. Z. (2015). Annonaceous acetogenins reverses drug resistance of human hepatocellular carcinoma BEL-7402/5-FU and HepG2/ADM cell lines. *International Journal of Clinical and Experimental Pathology*, 8, 11934.
- Rana, V. S. (2015). Fatty oil and fatty acid composition of *Annona squamosa* Linn. Seed kernels. *International Journal of Fruit Science*, 15, 79–84. <https://doi.org/10.1080/15538362.2014.931168>
- Raphael, E. C., Gideon, O. I., & Perpetua, N. U. (2010). Biochemical characteristics of the African nutmeg, *Monodora myristica*. *Agricultural Journal*, 5, 303–308. <https://doi.org/10.3923/aj.2010.303.308>
- Rocha, V., Duarte, M. C., Catarino, S., Duarte, I., & Romeiras, M. M. (2021). Cabo Verde's Poaceae Flora: A reservoir of crop wild relatives diversity for crop improvement. *Frontiers in Plant Science*, 12, Article 630217. <https://doi.org/10.3389/fpls.2021.630217>
- Romeiras, M. M., Essoh, A. P., Catarino, S., Silva, J., Lima, K., Varela, E., ... Duarte, M. P. (2023). Diversity and biological activities of medicinal plants of Santiago Island (Cabo Verde). *Heliyon*, 9, E14651. <https://doi.org/10.1016/j.heliyon.2023.e14651>
- Ruffo, C. K., Birnie, A., & Tenganas, B. (2002). Edible wild plants of Tanzania. *Technical Handbook*, 27, 766.
- Sapit, B., & Padulosi, S. (2012). On-farm conservation of neglected and underutilized crops in the face of climate change. In *On farm conservation of neglected and underutilized species: status, trends and novel approaches to cope with climate change: Proceedings of an International Conference, Frankfurt* (pp. 14–16).
- Sarpong, F. M., Amponsah, I. K., Armah, F. A., Jibira, Y., & Gyamfi, R. (2016). Anti-inflammatory and antioxidant activity of the ethanolic stem bark extract of *Pachypodanthium staudtii* Engl. & Diels (family Annonaceae). *World Journal of Pharmaceutical Sciences*, 4, 73–77.



- Scrimshaw, N. S. (2007). Book review: Handbook of nutraceuticals and functional foods. *Food and Nutrition Bulletin*, 28, 439. <https://doi.org/10.1177/156482650702800410>
- Shahriar, M., Khair, N. Z., Sheikh, Z., Chowdhury, S. F., Kamruzzaman, M., Bakhtiar, M. S. I., & Chisty, S. J. (2016). Characterization of phytoconstituents and potential bioactivity of *Annona reticulata* L. leaf extract. *Journal of Pharmacognosy and Phytochemistry*, 5, 42–45.
- Shaw, C. A., & Höglinger, G. U. (2008). Neurodegenerative diseases: Neurotoxins as sufficient etiologic agents? *Neuromolecular Medicine*, 10, 1–9. <https://doi.org/10.1007/s12017-007-8016-8>
- Sun, S., Liu, J., Kadouh, H., Sun, X., & Zhou, K. (2014). Three new anti-proliferative Annonaceous acetogenins with mono-tetrahydrofuran ring from graviola fruit (*Annona muricata*). *Bioorganic and Medicinal Chemistry Letters*, 24, 2773–2776. <https://doi.org/10.1016/j.bmcl.2014.03.099>
- Tamfu, A. N., Tagatsing Fotsing, M., Talla, E., Jabeen, A., Mbafor Tanyi, J., & Shaheen, F. (2021). Bioactive constituents from seeds of *Annona senegalensis* Persoon (Annonaceae). *Natural Product Research*, 35, 1746–1751. <https://doi.org/10.1080/14786419.2019.1634713>
- Tan, P. V., Boda, M., Enow-Orock, G. E., Etoa, F. X., & Bitolog, P. (2007). Acute and sub-acute toxicity profile of the aqueous stem bark extract *Enantia chlorantha* Oliver (Annonaceae) in laboratory animals. *Pharmacologyonline*, 1, 304–313.
- Tekuri, S., Pasupuleti, S., Konidala, K., & Pabbaraju, N. (2019). Pharmacological effects of *Polyalthia cerasoides* (Roxb.) Bedd.: A brief review. *Journal of Complementary Medicine Research*, 10, 38–49. <https://doi.org/10.5455/jcmr.20190108065022>
- Tolu-Odugbemi. (2008). *A textbook of medicinal plants from Nigeria*. Lagos: University of Lagos Press.
- Tsabang, N., Fokou, P. V. T., Tchokouaha, L. R. Y., Noguem, B., Bakarnga-Via, I., Nguepi, M. S. D., ... Boyom, F. F. (2012). Ethnopharmacological survey of Annonaceae medicinal plants used to treat malaria in four areas of Cameroon. *Journal of Ethnopharmacology*, 139, 171–180. <https://doi.org/10.1016/j.jep.2011.10.035>
- Umaru, H. A., Adamu, R., Dahiru, D., & Nadro, M. S. (2007). Levels of antinutritional factors in some wild edible fruits of northern Nigeria. *African Journal of Biotechnology*, 6, 1935–1938. <https://doi.org/10.5897/ajb2007.000-2294>
- Weenen, H., Nkunya, M. H. H., El-Fadl, A. A., Harkema, S., & Zwanenburg, B. (1990). Lucidene, a Bis(benzopyranyl) Sesquiterpene from *Uvaria lucida* ssp. *lucida*. *Journal of Organic Chemistry*, 55, 5107–5109. <https://doi.org/10.1021/jo00304a023>
- Weenen, H., Nkunya, M. H. H., Mgani, Q. A., Posthumus, M. A., Waibel, R., & Achenbach, H. (1991). Tanzanene, a Spiro Benzopyranyl Sesquiterpene from *Uvaria tanzaniae* Verdc. *Journal of Organic Chemistry*, 56, 5865–5867. <https://doi.org/10.1021/jo00020a030>
- Wertz, A. E., & Wynn, K. (2014). Selective social learning of plant edibility in 6- and 18-month-old infants. *Psychological Science*, 25, 874–882. <https://doi.org/10.1177/0956797613516145>
- Woguem, V., Fogang, H. P. D., Maggi, F., Tapondjou, L. A., Womeni, H. M., Quassinti, L., ... Barboni, L. (2014). Volatile oil from striped African pepper (*Xylopia parviflora*, Annonaceae) possesses notable chemopreventive, anti-inflammatory and antimicrobial potential. *Food Chemistry*, 149, 183–189. <https://doi.org/10.1016/j.foodchem.2013.10.093>
- Wu, T. Y., Yang, I. H., Tsai, Y. T., Wang, J. Y., Shiurba, R., Hsieh, T. J., ... Chang, W. C. (2012). Isodesacetylvaricin, an Annonaceous acetogenin, specifically inhibits gene expression of cyclooxygenase 2. *Journal of Natural Products*, 75, 572–576. <https://doi.org/10.1021/np200719r>
- Yanda-Shaba, E. (2017). Nutritive and anti-nutritive composition of *Xylopia parviflora*; seed obtained from Pati Shabakolo in Lavun local government area, Niger state, Nigeria. *Cell Biology*, 5, 53–56. <https://doi.org/10.11648/j.cb.20170505.12>
- Yanishlieva, N. V., & Marinova, E. M. (2001). Stabilisation of edible oils with natural antioxidants. *European Journal of Lipid Science and Technology*, 103, 752–767. [https://doi.org/10.1002/1438-9312\(200111\)103:11<752:AID-EJLT752>3.0.CO;2-0](https://doi.org/10.1002/1438-9312(200111)103:11<752:AID-EJLT752>3.0.CO;2-0)
- Yisa, J., Egila, J. N., & Darlinton, A. O. (2010). Chemical composition of *Annona senegalensis* from Nupe land, Nigeria. *African Journal of Biotechnology*, 9, 4106–4109.