Quantitative sidedress nitrogen recommendations for potatoes based upon crop nutritional indices


CIMO, Escola Superior Agrária; 5301-855 Bragança, Portugal

CECEA, University Trás-os-Montes e Alto Douro; 5001-911 Vila Real, Portugal

Abstract

The quantification of sidedress N rate from in-season information on crop N nutritional status is extremely important in order to use only the necessary amount of nutrient to achieve maximum yields without significant N losses from soil. During three consecutive years (1996, 1997 and 1998) field N fertilizer experiments with irrigated potato (Solanum tuberosum L., cv. Desirée) were conducted in Bragança (NE Portugal). Eight preplant treatments (0, 50, 100, 200 and 300 kg urea-N/ha and poultry manure, farmyard manure and municipal solid waste in rates equivalent to 100 kg organic-N/ha) were arranged as main plots and five sidedress N rates (0, 25, 50, 100 and 200 kg urea-N/ha) included as subplots. Petiole nitrate concentration, determined by a laboratory method (PNLab, g NO₃⁻/N kg⁻¹, dry weight basis) and by the portable RQflex reflectometer (PNRQflex, mg NO₃⁻/kg⁻¹, from fresh tissues), and leaf N content (LeafN, g kg⁻¹, dry weight basis) were used as N nutritional indices. From the five sidedress N rates applied over each of the preplant treatments, critical sidedress N rates were estimated for several different crop N nutritional status. This was achieved by establishing exponential asymptotic curves between sidedress N rates and tuber yields and solving the equations for 95% of maximum tuber yield. In a second step, multiple regression equations were established between the estimated critical sidedress N rates (SNrate, kg N/ha), as dependent variable, and each crop N nutritional indices (PNLab, PNRQflex or LeafN) and days after emergence (DAE). The equations obtained are:

SNrate = 182.7 − 4.146 × PNLab − 1.87 × DAE
SNrate = 161.0 − 0.013 × PNRQflex − 1.34 × DAE
SNrate = 562.2 − 8.416 × LeafN − 3.59 × DAE

These equations provide quantitative sidedress N rates for any level of crop N nutritional indices and sampling dates from 10 to 48 days after emergence.

Keywords: Solanum tuberosum; Crop N management; Decision support system; Plant analysis; Tissue testing

1. Introduction

Nitrogen is an important ecological factor. Its beneficial effects on crop growth are unquestionable, but if it
is used in excess could lead to environmental contami-
nation. Thus, the continuous improvement in fertiliser-
N recommendation programs is recognized worldwide.
The main limitation in several N recommendation sys-
tems all over the world is the estimation of soil contri-
bution to N nutrition of crops. In fact, reliable labora-
tory methods of soil analysis of universal acceptance
and widespread use, providing information concerning
soil N availability, are not yet developed. The problem
arises from the difficulty in preventing the time and rate
of mineralization of soil organic substrates (Dahnke
and Johnson, 1990).

The uncertainty of fertiliser-N recommendations
performed from soil analysis, before the start of the
growing season, and the high efficiency of N used at
sidedress (Westermann et al., 1988), have given im-
portance to fertilizer-N recommendation systems that
include information taken during the growing season.
Thus, an acceptable strategy for N management seems
to be the application at preplant of a part of N expected
to be needed with the necessary adjustments as side-
Plant analysis or tissue testing allow assessing the crop nu-
tritional status and, consequently, can be used to make
in-season N fertilization adjustments in order to better
balance N rates with crop needs.

The results of plant analysis are often interpreted by
critical levels or sufficiency ranges previously defined
for a given crop and for specific growing stages. The
critical level identifies the crop nutritional status, above
which there is no need to add more N and vice versa,
but not informs about the amount of nutrient to apply.

The nitrogen nutrition index (NNI), which could diagnose the intensity
of N deficiency, as well as situations of luxury con-
sumption. The NNI can be determined, for any crop and
at any time, as the ratio between the actual plant N% and the critical plant N% corresponding to adequate
N uptake. Nitrogen nutrition index values close to
1 identify situations of non-limiting N supply. Values
greater than 1 indicate luxury consumption. The closer
the value is to 0, the greater the intensity of the N de-

2. Material and methods

2.1. Field experiments and laboratory analysis

The survey was conducted between 1996 and
1998 in North-eastern potato-growing region of Por-
tugal (41°N latitude and 6°W longitude), with a
Mediterranean-type climate, hot and dry during the
summer growing season. Irrigation is a must for suc-
cessful potato growing. The soil is a eutric Cambisol,
loamy textured, with pH(H2O) of 6.5 and organic mat-
ter content of 15 g kg⁻¹. Extractable P and K (am-
monium lactate and acetic acid) ranged from 20 to
49 mg kg⁻¹ and 84 to 149 mg kg⁻¹, respectively. Pre-
vious crops were corn for silage in summer followed
by triticale for silage, grown as winter inter-crop.

The field experiments were arranged as a split-plot
design. The main plots consisted on the following eight
preplant N treatments: 0, 50, 100, 200 and 300 kg urea-
N/ha and poultry manure, farmyard manure and a mu-
nicipal solid waste, applied in variable rates corre-
sponding to 100 kg organic-N/ha. The main plots were
split into subplots to include five sidedress N rates (0,
25, 50, 100 and 200 kg urea-N/ha). In 1996 and 1997,
the sidedress treatment of 200 kg N/ha was not applied.
over the preplant treatments of 200 and 300 kg N/ha. The plots were laid out through the field in three blocks (three replicates). Each individual subplot consisted of 5 m long seven rows.

Phosphorus and potassium rates applied in 1996, 1997 and 1998 were, respectively, 26, 52 and 26 kg P/ha and 75, 125 and 66 kg K/ha. The urea of the preplant treatments, the organic amendments and P (superphosphate, 18% P₂O₅) and K (potassium chloride, 60% K₂O) were broadcast and incorporated with preplanting tillage. Whole pre-sprouting tubers with sizes of 28–45 mm (1996) and 45–60 mm (1997 and 1998) were machine-planted on 28 May 1996, 28 May 1997 and 21 May 1998. Seed tubers were planted with a row spacing of 0.70 m, and in-row spacing of 0.32 m. Crop emergence (50% of visible shoots) occurred 14, 13 and 15 days after planting in 1996, 1997 and 1998, respectively. The urea of the sidedress N treatments was applied onto soil surface on 17 July 1996, 9 July 1997 and 24 June 1998 and incorporated with overhead irrigation water. Depth and interval irrigation application were based on FAO methods (Doorenbos and Pruit, 1977). Crop protection included weeds, late blight (Phytophthora infestans) and beetle (Leptinotarsa decemlineata) control. Detailed information of pesticides, rates and dates of application were previously reported (Rodrigues et al., 2001). The harvests occurred in late September after the full senescence of the plants. The fresh weights of marketable tubers (>35 mm) were recorded.

Fifty to sixty of the most recently matured leaves were taken from each plot. One-third of the full leaves were used for leaf N analysis. The remaining leaves were separated into blades and petioles for petiole nitrate analysis. The full leaves' samples and half of the petioles' samples were oven-dried (65 °C) and ground. Leaf N was determined by Kjeldahl method, in a Kjeltec Auto 1030 Analyzer. From dried petiole subsamples, nitrate analysis were performed by adding 50 ml of distilled water to 1.0 g of dry sample. The mixtures were shaken for 1 h and filtered. Nitrate-N concentrations were determined by the sulfanilamine method (Houba et al., 1989) and analysed in a segmented flow analyser. For quick testing, 5 g of the fresh petioles were macerated and boiled for 15 min in 100 ml of distilled water. After cooling, final volumes of 300 ml were lined up and nitrate concentrations in extracts determined by Reflectoquant test strips, read with the RQflex reflectometer (MERCK). During the 3 years survey, between 10 and 48 days after emergence, nine different sampling dates were chosen.

2.2. Sidedress N rates quantification as a function of crop N nutritional status and sampling dates

The sidedress N rates quantification as a function of crop N nutritional status and sampling dates involved two distinct steps. In the first one, we determined the critical sidedress N rates. By analogy with critical nutrient concentration in a given tissue, we define critical sidedress N rate as the point, above which the crop does not respond to more nutrient applied at sidedress. In practical terms, we determine the critical sidedress N rates after establishing the relationships between applied sidedress N rates and tuber yields and solving the equations for 95% of maximum tuber yield. This is an environmentally sound approach, since we accepted a yield reduction of 5% as is usual in plant analysis studies. As was previously described, each of the preplant plots was sidedressed with five N doses, arranged as subplots. Thus, the response of the crop to N applied as sidedress allowed estimating critical sidedress N rates from very different crop N nutritional status, arising from the different urea-N and organic-N preplant treatments.

The relationships between the sidedress N rates and tuber yields were fitted by exponential asymptotic equations of the type of

\[ y = a - b \exp(c x) \]

where \( y \) is tuber yield (Mg/ha), \( x \) is applied sidedress N (kg/ha) and \( a \) and \( b \) are coefficients estimated from experimental data. The exponential coefficient \( c \) was predetermined to avoid non-linear regression analysis, which needs much computing time, as suggested by Neeteson and Wadman (1987). The exponential coefficient \( c \) was estimated by:

\[ c = \frac{d \ln(d)}{N_{\text{max}}} \]

where \( d \) has values of 0.03, 0.05, 0.10, 0.20, 0.35, 0.60, 0.90, 1.50 and 2.79 with \( N_{\text{max}} \) being the highest level of fertiliser N applied in kg/ha. Thus, the exponential coefficient could assume several values to get flexibility of the curves. The \( c \) value, which yielded the
lowest residual sum of squares, defines the curve of best fit. As above-mentioned, the critical sidedress N rates were obtained solving these equations for 95% of maximum tuber yield. Considering the continuous increase of this type of curves, maximum tuber yield is the point corresponding to the higher sidedress N rate applied. However, when 300 kg N/ha were applied at preplant, as well as 200 kg N/ha in 1997 and 1998, the effect on tuber yield of sidedress N was not evident. In 1997, when 200 and 300 kg N/ha were applied at preplant, the exponential curves showed the concavity turned upwards. In 1996 with 300 kg N/ha and in 1998 with 200 kg N/ha, the curves showed a continuous decrease in tuber yield starting at the control treatment. In 1998, when 300 kg N/ha were applied at preplant, the slight increase observed in tuber yield with sidedress N was less than 5% of maximum tuber yield. Thus, in all these five situations, we considered the critical sidedress N rate as 0 kg N/ha.

In a second step, multiple regression equations were proposed to obtain quantitative sidedress N rates as a function of crop N nutritional indices and days after emergence. The multiple regression equations were established in the form of:

\[ z = a + by + cx \]

where \( z \) is recommended sidedress N rate, \( y \) is a given nitrogen nutritional index, \( x \) is days after crop emergence and \( a, b \) and \( c \) are coefficients determined from experimental data. The equations take into account the decreasing linear relationship observed between the N nutritional indices and the critical sidedress N rates as calculated in the first step. They also consider the linear decrease on the petiole nitrate concentration and leaf N throughout the growing season, observed between 10 and 48 days after emergence, independently of the preplant treatment.

3. Results

The tuber yield responses to the N applied at preplant and sidedress are shown in Fig. 1. The yield responses to preplant N were plotted on the \( y \) lines on each of the individual small figures, corresponding to the plots where no N was applied as sidedress. The Fig. 1 also includes the fitting curves corresponding to the exponential asymptotic model and their equations represent the tuber yield responses to sidedress N rate. Minimum tuber yields were recorded on control treatments, where N was neither applied at preplant nor sidedress (Fig. 1, Pp0). Tuber yields recorded on control plots were 42.8, 23.5 and 32.3 Mg/ha in 1996, 1997 and 1998, respectively. As higher the N rate applied at preplant, the lower the tuber yield response to sidedress N and vice versa. Maximum tuber yields close to 50 Mg/ha, resulting from diverse combinations of preplant and sidedress N rates, were recorded for all the years. Thus, this value could define the yield potential of the agro-system.

The estimated critical sidedress N rates ranged from 0 to 149 kg N/ha (Table 1). Nought and lower values were found on plots where high amounts of N were applied at preplant. The preplant treatments, which do not provide enough N for adequate crop growth, such as control, 50 kg urea-N/ha and the treatments consisting on organic amendments as nitrogen source, lead to higher values of critical sidedress N rates.

Quantified sidedress N rates as a function of petiole nitrate concentrations and sampling dates are given by the multiple regression equation presented in Fig. 2. The negative partial regression coefficient (−4.15) for the PN Lab variable, indicates lower recommended sidedress N rates as higher petiole nitrate concentration is for a given sampling date. For a given value of petiole nitrate concentration, the recommended sidedress N rate decreases as the growing season progresses, since the partial regression coefficient for DAE is also negative (−1.87). The potential and the functioning of the

<table>
<thead>
<tr>
<th>Preplant treatments (kg N/ha)</th>
<th>1996</th>
<th>1997</th>
<th>1998</th>
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<tbody>
<tr>
<td>0 (control)</td>
<td>92</td>
<td>149</td>
<td>80</td>
</tr>
<tr>
<td>50 (urea)</td>
<td>62</td>
<td>120</td>
<td>73</td>
</tr>
<tr>
<td>100 (urea)</td>
<td>58</td>
<td>95</td>
<td>32</td>
</tr>
<tr>
<td>200 (urea)</td>
<td>24</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>300 (urea)</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>100 (poultry manure)</td>
<td>57</td>
<td>136</td>
<td>121</td>
</tr>
<tr>
<td>100 (farmyard manure)</td>
<td>121</td>
<td>144</td>
<td>69</td>
</tr>
<tr>
<td>100 (man. sol. waste)</td>
<td>68</td>
<td>138</td>
<td>89</td>
</tr>
</tbody>
</table>

The values were obtained solving the equations of Fig. 1 for 95% of maximum tuber yield.

Table 1

Critical sidedress N rates estimated from tuber yield response to nitrogen applied as sidedress over each preplant treatment of the 3 years of study.
Fig. 1. Relationships between applied sidedress N (x) and tuber yield (in 1996, \(y_{96}\); 1997, \(y_{97}\); 1998, \(y_{98}\)). Pp0, Pp50, Pp100, Pp200, Pp300, PpPM, PpFM and PpMSW are the preplant treatments, which consisted on 0, 50, 100 and 200 kg urea-N/ha and 100 kg organic-N/ha as poultry manure, farmyard manure and municipal solid waste, respectively. Vertical bars are standard deviations of the means.

The equation could be fully understood following this example: if at 20 days after emergence, the petiole nitrate-N concentrations is 30 g kg\(^{-1}\), the recommended sidedress N dose is 21 kg/ha; if at the same sampling date, the petiole nitrate-N concentration is only 10 g kg\(^{-1}\), the estimated sidedress N dose is 104 kg/ha, and if the petiole nitrate-N concentration is 10 g kg\(^{-1}\) at 45 DAE, the sidedress N required is 57 kg/ha.
Quantified sidedress N rates as a function of petiole nitrate concentrations (performed in fresh tissues with the RQflex reflectometer) and days after emergence can be obtained from the regression equation shown in Fig. 3. The pattern of the response surface is similar to that of Fig. 2, notwithstanding the fact that the partial regression coefficient for PNRQflex (−0.013) is much different due to the units in which the results are expressed.

Taking into consideration the simplicity of the portable tool, its accuracy was double checked using standard Ca(NO₃)₂ solutions and their results compared with the laboratorial analysis. We stress that RQflex readings and laboratory analysis were performed on random sub-samples of the same petiole sample. The relationship between nitrate concentrations in Ca(NO₃)₂ solutions and RQflex readings is shown in Fig. 4. The relationship between petiole nitrate concentrations determined from laboratory analysis and with the RQflex reflectometer is shown in Fig. 5. The slope of the linear regression between nitrate concentrations in Ca(NO₃)₂ solutions and RQflex readings is 1.03 (Fig. 4). Testing the hypotheses $H₀: β₁ = 1$ versus $H₁: β₁ ≠ 1$, the null hypothesis was not rejected ($p < 0.01$), meaning that RQflex readings do not need calibration in the reactivity range of reflectoquant test strips. It was also attained a close correlation ($r = 0.96$) between petiole nitrate concentrations determined by the laboratory method and RQflex reflectometer (Fig. 5), even considering the dissimilar process of extracts preparation. For laboratory analysis, we used dried and ground petioles, whilst the extracts were prepared from fresh tissues for RQflex readings.

The multiple regression equation providing the recommended sidedress N rates as a function of leaf N and DAE is shown in Fig. 6. The multiple coefficient of determination ($R^2 = 0.59$) is not much different to that obtained with the petiole nitrate concentration, despite the latter being the most common N nutritional status index in potato crop. As with petiole nitrate concentration, as higher the leaf N content is, the lower is the recommended sidedress N rate for a given sampling date. For similar levels of leaf N content, the recommended sidedress N rates also decreases with time. Fig. 7 shows the scatter diagram of the multiple regression equations residues. The deviation of observed to estimated values by regression surface ranged from $−57$ to $+67$, $−61$ to $+54$ and $−56$ to $+59$ kg N/ha, re-
spectively for PNLab, PNRQflex and LeafN. The dots were plotted by sampling date to allow for a follow up of their evolution during the growing season. We could observe some variation between sampling dates of the different years. However, the pattern of residues distribution throughout the season seems to support the model application between 10 and 48 days after crop emergence.

4. Discussion

The exponential asymptotic model fitted very well to the data in the range of sidedress N rates used, because there were no evident decreases in tuber yields for the higher N rates. It is common in potato crop the development of an ample plateau where tuber yield
Fig. 6. Plane and respective equation, which provide the recommended sidedress N rate (SRate) as a function of leaf N (LeafN) and days after crop emergence (DAE).

Tuber yield was dependent to preplant and sidedress N rate and to the year. The effect of applied N on tuber yield is well documented and has been reported worldwide. The yearly differences found on tuber yield, mainly in control plots, could be justified by differences in N availability from soil reserves. An in situ incubation technique, carried out in the context of this study, showed significant differences in soil inorganic N released during the growing season (Rodrigues, 2004). Neeteson and Wadman (1987) reported a clarifying result on this topic. Using data of 99 field experiments scattered over the Netherlands, the authors found economically optimum N rates ranging between 0 and 400 kg/ha. Differences in soil N availability were considered as the main factor influencing the results. As expected, the estimated critical sidedress N rates were higher, as lower the preplant N rates and vice versa. Thus, crop response to sidedress N rate depends on N release from soil reserves and N applied at preplant.

Singh (1993) reported optimal sidedress N rates decreasing as higher the petiole nitrate-N concentrations. From petiolar NO₃-N concentrations of 0.5, 1.0, 1.5, 2.0 and 2.5% at 30 days after planting, Singh (1993) obtained the corresponding optimal rates of sidedress N of 142, 116, 90, 64 and 37 kg/ha for cv. Kufri Chandramukhi and 183, 164, 146, 127 and 108 kg/ha for cv. Kufri Badshah. If the multiple regression equation of the Fig. 2 is running with the above petiolar NO₃-N values and considering 30 days after planting similar to 15 days after emergence, the sidedress N rates recommended are 134, 113, 93, 72 and 51 kg/ha. Singh (1993) and our results could not be considered much dissimilar, notwithstanding the differences in experimental conditions in India and Portugal and also the cultivars.

There is abundant literature reporting critical levels for petiole nitrate concentration in potatoes. The critical level represents the crop nutritional status, above which the probability of crop response to more N is low. Thus, if the equation is solved using critical petiole nitrate concentrations, the sidedress N rate recommended would be close to 0 kg/ha. Westcott et al.
Fig. 7. Residues of recommended sidedress N rates obtained as the difference between observed and estimated values from the equations of Figs. 2, 3 and 6. PNLab, data of petiole nitrate concentration (dry weight basis); PNRQflex, petiole nitrate concentration (fresh weight basis); LeafN, leaf N concentration. Each vertical line of dots represents the residues of the different sampling dates. The open rings represent the mean values of residues of each sampling date.

(1991) reported critical petiole nitrate levels in potato (cv. Russet Burbank) of 25 g kg\(^{-1}\) at tuber initiation (TI), 14 g kg\(^{-1}\) at TI + 21 days and 10 g kg\(^{-1}\) at TI + 42 days. Considering TI equivalent to our time scale of 20 DAE, the equation of the Fig. 2 gives sidedress N rates of 42, 48 and 25 kg/ha, respectively. If the equation is solved with other published critical levels (e.g. Porter and Sisson, 1991; Singh, 1993; Bélanger et al., 2003), estimated sidedress N rates are always slightly higher than zero. This apparent overestimation of N rate above 0 kg/ha could be justified, taking into account the manner in which critical levels are determined. The critical levels reported in literature were determined by different ways: fitting the curves between petiole nitrate concentrations and tuber yields (Gupta and Saxena, 1976; Singh, 1993; Westcott et al., 1991, 1993; Sims et al., 1995). However, we are unaware of any work using this specific equipment, but from these results, it seems to show a high usefulness potential. The farmers and field consultants need simple equipments, like RQflex reflectometer, to diagnose the N nutritional status of annual crops. The growth rate of annual crops is high and laboratories often take too much time to provide a diagnostic. So, if the nutritional status of a crop approaches a deficiency level at the time of sampling, a high yield loss could occur when corrective measures are delayed.

Leaf N is not commonly used as N nutritional index in potatoes. The exception being situations where a multi-elemental analysis, to monitor the accuracy of a fertilizer recommendation program, is desired. The petiole nitrate concentration is most widespread when the purpose is the early diagnostic of a N deficiency, to allow for in-season adjustments of N doses. However, leaf N can also be used as N nutritional index. Eventylo (1989) reported critical leaf N with cv. Superior of 58 g kg\(^{-1}\) at 14 days after emergence and 51.6 g kg\(^{-1}\) at midbloom (≈35 DAE). Using such values to solve Fig. 6 equation, one obtains 24 and 2 kg/ha, respectively, as recommended sidedress N rates. These values are very close to zero, the theoretically expected result and show agreement between the methodologies reported in this work and the classical approach by critical levels.

The strategy proposed in this work consists on transferring of at least part of the decision of N quantification from preplant to sidedress time. It assumes the difficulties on N rate quantification before the start of
the growing season found worldwide in many N recommendation programs. We suggest the application of moderate N rates at preplant with final corrections of N fertilisation at sidedress, based upon N nutritional indices. The plots of residues (Fig. 7) show that we could use the equations of Figs. 2, 3 and 6 between 10 and 48 days after crop emergence. This period covers the most adequate part of growing season to correct a nitrogen shortage. Delaying fertiliser-N application may reduce the N use efficiency.

Acknowledgements

The authors thank Rita Diz and Ana Pinto for laboratory assistance, Sergio Deusdado for helping on statistical software and Chrys Chrystello for English improvement. Financial support was provided by the Ministry of Agriculture, Rural Development and Fisheries through the Project PAMAF 6107, and FCT through the Project PRAXIS P/AGR/11045/98.

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